



**OsteoBiol®**  
by Tecnoss

Collagen-based biomaterials

REGENERATION SCIENCE

INSPIRED BY NATURE



# Index

<b>Our mission</b>	4
<b>The devices</b>	
<b>Collagenic bone matrix</b>	9
<i>GTO®</i>	14
<i>mp3®</i>	15
<i>Gel 40</i>	16
<i>Putty</i>	17
<i>Gen-Os®</i>	18
<i>Apatos®</i>	19
<i>TSV Gel</i>	20
<i>Sp-Block</i>	21
<i>Lamina®</i>	22
<b>Collagen membranes</b>	23
<i>Evolution</i>	28
<i>Derma</i>	29
<b>The clinical experience</b>	30
<i>GTO®</i>	32
<i>mp3®</i>	38
<i>Gel 40</i>	44
<i>Putty</i>	48
<i>Gen-Os®</i>	52
<i>Apatos®</i>	58
<i>Sp-Block</i>	62
<i>Lamina®</i>	66
<i>Evolution</i>	72
<i>Derma</i>	78
<b>Our history</b>	84
<b>Quality system</b>	85
<b>Distribution network</b>	86
<b>Our international courses</b>	88
<b>Bone, Biomaterials &amp; Beyond Academy</b>	89
<b>OsteoBiol® product codes</b>	90



# OUR MISSION

«To create xenogenic bone substitutes and collagen membranes as similar as possible to autogenous bone and soft tissues.»

**Giuseppe Oliva, MD**

Founder of **Tecnoss S.r.l.** & inventor of **OsteoBiol®** biomaterials

# OUR VISION

We want to reduce the invasiveness of surgical procedures and improve patients' lives by delivering innovative and scientifically validated xenogenic collagenated biomaterials to healthcare professionals.

# **PATIENTS FIRST**

Combining the best skills and the best materials, within the limits and guidelines provided by scientific evidence, is the key for clinical success: however, let us all remember that the patients are and will always be the centre of all our attentions.

Meeting their expectations, helping them to recover function and esthetics with long term success<sup>(1,2)</sup> is the greatest reward for any surgeon and fulfillment of our company mission.

(1) Barone A et al. J Periodontol, 2011 Feb; 82(2):219-26

(2) Scarano A et al. J Oral Maxillofac Surg, 2010 Aug;68(8):1869-73



# Tecnoss®

## Tecnoss®

The Manufacturer

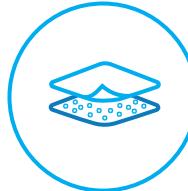
### OsteoBiol®

by Tecnoss

The Biomaterials line



Collagenic  
bone matrix



Collagen  
membranes

The Devices

# **COLLAGENIC BONE MATRIX**



# THE ADVANTAGES OF A

## WHY COLLAGEN?

Natural human hard bone tissue contains approximately 25% collagen. Collagen is also considered the ideal substrate for bone-forming cells as it is a chemotactic molecule<sup>(1,2)</sup> and stimulates new blood vessels formation<sup>(3)</sup>.

## SIMILARITY TO AUTOGENOUS BONE

About 22% collagen is retained in each OsteoBiol® xenogenic granule, making it chemically and physically similar to autogenous bone<sup>(4)</sup>.

## MESENCHYMAL STEM CELLS DIFFERENTIATION

Collagen acts as a chemoattractant towards mesenchymal stem cells and promotes their differentiation into osteogenic cells<sup>(3)</sup>.

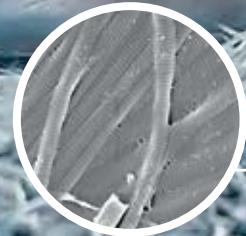
## NEW BONE FORMATION

Collagen also recruits osteogenic cells such as osteoclasts and osteoblasts at the defect site. The former progressively resorb OsteoBiol® collagenic xenograft<sup>(1,2)</sup> while the latter simultaneously deposit newly formed bone<sup>(1,2)</sup>.



Osteoblasts landed on OsteoBiol® Dual-Phase bone matrix.

Author: Prof Ulf Nannmark, University of Göteborg, Sweden



Collagen fibres embedded within OsteoBiol® collagenic xenografts.

Author: Prof Ulf Nannmark, University of Göteborg, Sweden

# COLLAGENIC XENOGRAFT

OsteoBiol® Dual-Phase bone granules are not ceramized<sup>(4)</sup> and their gradual and progressive resorption leads to an adequate amount of newly formed bone at re-entry time.

Osteogenic cells live thanks to the blood supply, which is essential for the success of any bone regeneration procedure. An *in vitro* study demonstrated that OsteoBiol® Dual-Phase bone substitutes promote new blood vessel formation<sup>(3,5)</sup>, thus providing osteoblasts and osteoclasts with oxygen and nutrients.

The bloodstream also provides growth factors, a set of molecules that play a key role in bone regeneration, including Vascular Endothelial Growth Factor (VEGF) a crucial protein for neovascularization and angiogenesis.

Due to its high similarity with the gold standard for bone regeneration<sup>(4)</sup>, the OsteoBiol® collagenic product line is a valid alternative to autogenous bone for various surgical procedures<sup>(6-11)</sup>.

GRADUAL  
RESORPTION

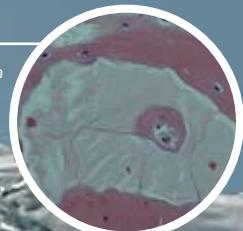
BLOOD VESSELS

GROWTH FACTORS

AN ALTERNATIVE  
TO GOLD STANDARD

Newly formed bone surrounding a GTO® collagenic granule.

Author: Prof Ulf Nannmark, University of Göteborg, Sweden



1. Nannmark U et al. Clin Implant Dent Relat Res, 2008 Dec;10(4):264-70
2. Falacho RI et al. Molecules, 2021 Mar 2;26(5):1339
3. Rombouts C et al. Dent Mater J, 2016 Dec 1;35(6):900-907
4. Figueiredo M et al. J Biomed Mater Res B Appl Biomater, 2010 Feb;92(2):409-419
5. Jeanneau C et al. Materials, 2024; 17(3):625
6. Correia F et al. Materials (Basel), 2021 Jun 21;14(12):3439
7. Correia F et al. Materials, 2023; 16(3):1220
8. Correia F et al. Dentistry Journal, 2024; 12(2):33
9. Felice P et al. Int J Periodontics Restorative Dent, 2017 Jul/Aug;37(4):469-480
10. Villa G et al. Int J Periodontics Restorative Dent, 2023 July-Aug;43(4):435-441
11. Happe A et al. J Clin Med, 2023 Nov 9;12(22):7013

# THE ADVANTAGES OF A

## OSTEOBIOL® COLLAGENIC XENOGRAFT: A UNIQUE PRODUCT LINE

Tecnoss® developed several biomaterials for every clinical indication<sup>(1)</sup> to provide the best handling, the ideal granulometry and consistency, and optimal regenerative results within adequate re-entry time.

Today it is finally possible to achieve predictable clinical success in most regenerative protocols without the availability limitations of autogenous bone<sup>(1-5)</sup>.

Enjoy one of the widest and most complete product line of biomaterials, with the security and support of more than 25 years of clinical research.



Predictable results in sinus lift procedures



Easy-handling collagenated xenograft



Predictable outcomes in the esthetic area

# DUAL-PHASE BIOMATERIAL



Various clinical studies showed that the OsteoBiol® product line can contribute to the formation of an appropriate amount of newly formed bone<sup>[6,7]</sup>. Hence, implants are stable over long follow-up periods (> 5 years)<sup>[8-11]</sup> with a minimal bone loss around the implant surface.

Hard and soft tissue volume loss is a common post-operative complication. Scientific studies report that OsteoBiol® collagenic xenografts prevent both hard<sup>[12]</sup> and soft<sup>[13]</sup> tissue volume shrinkage.

Since the last 25 years, scientific studies showed that OsteoBiol® product line is reliable and safe over long follow-up periods<sup>[8-11]</sup> and in complex regenerative procedures<sup>[4,14]</sup>.

25 years of clinical experience<sup>[1]</sup> demonstrated that the OsteoBiol® bone substitutes are a valid alternative to autogenous bone particles in several surgical procedures<sup>[2-5]</sup>.

IMPLANT STABILITY  
OVER LONG PERIODS

PREVENTION OF  
VOLUME LOSS

SAFETY

A LONG HISTORY OF  
CLINICAL EXCELLENCE

1. Romasco T et al. *J Funct Biomater*, 2022 Aug;13(3):121
2. Felice P et al. *Int J Periodontics Restorative Dent*, 2017 Jul/Aug;37(4):469-480
3. Correia F et al. *Dentistry Journal*, 2024; 12(2):33
4. Villa G et al. *Int J Periodontics Restorative Dent*, 2023 July-Aug;43(4):435-441
5. Happe A et al. *J Clin Med*, 2023 Nov 9;12(22):7013
6. Hirota A et al. *Oral Maxillofac Surg*, 2020 Sep;24(3):299-308
7. Barone A et al. *J Periodontol*, 2008 Aug;79(8):1370-7
8. Pistilli R. et al. *J Dent*, 2022 Jun;121:104137
9. Testori T et al. *Int J Oral Implantol (Berl)*, 2024 May 27;17(2):189-198
10. Schuh PL et al. *Materials (Basel)*, 2021 Sep 9;14(18):5180
11. Esposito M. et al. *Clinical Trials In Dentistry*, 2020;02(4):21-37
12. Barone A. et al. *Clin Implant Dent Relat Res*, 2017 Aug;19(4):750-759
13. Fischer KR et al. *Clin Implant Dent Relat Res*, 2019 Oct;21(5):923-930
14. Köttgen C et al. *Int J Esthet Dent*, 2024 May 10;19(2):152-169



## Porcine origin

Codes	Packaging	Size	Granulometry	Mean observed re-entry time
-------	-----------	------	--------------	-----------------------------

MU0005S	1 Syringe	0,5 cc	0,6-1 mm	About 5 months
MU0020S	1 Wide-tip syringe	2 cc	0,6-1 mm	About 5 months

## Equine origin

Codes	Packaging	Size	Granulometry	Mean observed re-entry time
-------	-----------	------	--------------	-----------------------------

MU0005E	1 Syringe	0,5 cc	0,6-1 mm	About 5 months
MU0020E	1 Wide-tip syringe	2 cc	0,6-1 mm	About 5 months

## Description

Collagenated bone mix granules pre-hydrated blended with a thermosensitive copolymer

Pre-hydrated granules and TSV Gel

Heterologous bone mix

## Characteristics

**GTO®** is sticky, gradually resorbable and osteoconductive<sup>(1)</sup>.

Thanks to its high collagen content<sup>(1)</sup>, **GTO®** allows an excellent rate of new bone formation<sup>(2-4)</sup>, delivering adequate graft volume preservation<sup>(5)</sup>, healthy new bone tissue and, ultimately, a successful implant rehabilitation<sup>(2-7)</sup>. **GTO®** is a mixture of heterologous collagenated cortico-cancellous granules and **TSV Gel**, the component that provides its sticky properties<sup>(8)</sup>.

## How to handle GTO®

**GTO®** has been conceived as a universal biomaterial, easily adaptable to any bone defect.

It is ready-to-use, pre-hydrated and remains stable into the defect site.

Thus, clinicians can skip the hydration step with saline or blood, saving time and decreasing the risk of accidental exposure to pathogens.



1. Jeanneau C et al. Materials, 2024; 17(3):625
2. Canullo L et al. J Dent, 2024 Nov;150:105337
3. Passarelli PC et al. Am J Dent, 2024 Sept;37(SIA):9A-12A
4. Lopez MA et al. Am J Dent, 2024 Sept;37(SIA):41A-44A
5. Menini M et al. Dentistry Journal, 2024; 12(7):198
6. Passarelli PC et al. Am J Dent, 2024 Sept;37(SIA):4A-8A
7. Cinquini C et al. Clin Implant Dent Relat Res, 2025 Aug;27(4):e70085
8. Taniguchi Z et al. Int J Oral Maxillofac Implants, 2024 Oct 4;0(0):1-28



## Porcine origin

Codes	Packaging	Size	Granulometry	Mean observed re-entry time
A3095FS	1 Syringe	0,5 cc	0,6-1 mm	About 5 months
A3005FS	1 Syringe	1 cc	0,6-1 mm	About 5 months
A3010FS	1 Wide-tip syringe	2 cc	0,6-1 mm	About 5 months
A3210FS	1 Wide-tip syringe	2 cc	1-2 mm	About 5 months

## Equine origin

Codes	Packaging	Size	Granulometry	Mean observed re-entry time
A3095FE	1 Syringe	0,5 cc	0,6-1 mm	About 5 months
A3005FE	1 Syringe	1 cc	0,6-1 mm	About 5 months
A3010FE	1 Wide-tip syringe	2 cc	0,6-1 mm	About 5 months
A3210FE	1 Wide-tip syringe	2 cc	1-2 mm	About 5 months

## Description

Collagenated bone granules pre-hydrated with collagen gel  
Heterologous cortico-cancellous bone mix

## Characteristics

**mp3®** is a Dual-Phase pre-hydrated biomaterial. Hence, it is possible to skip the hydration phase and decrease the risk of accidental exposure of the material to pathogens during the manipulation phase.

Dual-Phase collagenic bone granules are endowed with characteristics very similar to autogenous bone<sup>(1)</sup>, and can be used as a viable alternative to autogenous bone providing similar performances over time<sup>(2)</sup>.

**mp3®** granules are gradually resorbable<sup>(3,4)</sup>, and preserve the original shape and volume of the bone surrounding the defect<sup>(5)</sup> allowing a successful regeneration.

## How to handle mp3®

**mp3®** must be used in a sterile environment.

After adapting the material to the defect shape, it is necessary to remove non stable residues before proceeding to the soft tissues suture.



1. Correia et al. Materials (Basel), 2021 Jun 21;14(12):3439
2. Correia et al. Dentistry Journal, 2024; 12(2):33
3. Nannmark U et al. Clin Implant Dent Relat Res, 2008 Dec; 10(4):264-70
4. Giuliani A et al. Clin Oral Investig, 2018 Jan;22(1):505-513
5. Barone A et al. Clin Oral Implants Res, 2016 Nov;27(11):e105-e115



## Porcine origin

Codes	Packaging	Size	Granulometry	Mean observed re-entry time
-------	-----------	------	--------------	-----------------------------

05GEL40S	1 Syringe	0,5 cc	≤ 0,300 mm	About 4 months
----------	-----------	--------	------------	----------------

### Description

Collagenic bone matrix and collagen gel  
Heterologous cortico-cancellous bone mix

## Characteristics

The composition of **Gel 40** facilitates the gentle detachment of the sinus membrane<sup>(1)</sup>.

Cortico-cancellous Dual-Phase bone granules provide the necessary scaffold function<sup>(2)</sup>, allowing the subsequent invasion of regenerative cells and new bone formation<sup>(1,2)</sup>.

**Gel 40** provides an osteoconductive matrix for new bone growth, while promoting soft tissue healing, when used in narrow self-contained bone defects<sup>(1,2)</sup>.

## How to handle Gel 40

The distinctive viscosity and density of **Gel 40** facilitate the handling of the bone gel.

It was documented that, in case of adequate height of the bone crest, immediate implant placement reduces the physiological shrinkage of the biomaterial<sup>(1)</sup> after crestal access sinus lifting procedures.



1. Lombardi T et al. Int J Implant Dent, 2022 Jul 22;8(1):32

2. Kobe T et al. Clin Exp Dent Res, 2024 Feb;10(1):e853



## Porcine origin

Codes	Packaging	Size	Granulometry	Mean observed re-entry time
HPT52S	1 Syringe	0,25 cc	$\leq 0,300$ mm	About 4 months
HPT09S	1 Syringe	0,5 cc	$\leq 0,300$ mm	About 4 months
HPT61S	1 Wide-tip syringe	1 cc	$\leq 0,300$ mm	About 4 months

## Equine origin

Codes	Packaging	Size	Granulometry	Mean observed re-entry time
HPT52E	1 Syringe	0,25 cc	$\leq 0,300$ mm	About 4 months
HPT09E	1 Syringe	0,5 cc	$\leq 0,300$ mm	About 4 months
HPT61E	1 Wide-tip syringe	1 cc	$\leq 0,300$ mm	About 4 months

## Description

Collagenic bone paste

Heterologous cortico-cancellous bone mix

## Characteristics

**Putty** is a pre-hydrated injectable bone paste containing collagenic bone granules with a granulometry of  $\leq 300$  mm.

The small diameter bone granules facilitates the application of **Putty** into peri-implant lesions with intact bony walls<sup>(1)</sup>.

**Putty** should be used in defects able to firmly contain it<sup>(1-3)</sup>, in order to stabilize the graft and allow its gradual resorption and new bone deposition.

## How to handle Putty

**Putty** should be injected into the defect and adapted to its morphology without extensive compression.

Any non-stable residue must be removed before soft tissue suture.



1. Barone A et al. Eur J Implant Prosthodontics, 2006, 2:99-106
2. Cassetta M et al. Int J Periodontics Restorative Dent, 2012 Oct;32(5):581-9
3. Saglanmak A et al. J Clin Med, 2024 Apr 11;13(8):2225



## Porcine origin

Codes	Packaging	Size	Granulometry	Mean observed re-entry time
M1052FS	1 Vial	0,25 g	0,25-1 mm	About 5 months
M1005FS	1 Vial	0,5 g	0,25-1 mm	About 5 months
M1010FS	1 Vial	1 g	0,25-1 mm	About 5 months
M1020FS	1 Vial	2 g	0,25-1 mm	About 5 months
M0210FS	1 Vial	1 g	1-2 mm	About 5 months
M0220FS	1 Vial	2 g	1-2 mm	About 5 months

## Equine origin

Codes	Packaging	Size	Granulometry	Mean observed re-entry time
M1052FE	1 Vial	0,25 g	0,25-1 mm	About 5 months
M1005FE	1 Vial	0,5 g	0,25-1 mm	About 5 months
M1010FE	1 Vial	1 g	0,25-1 mm	About 5 months
M1020FE	1 Vial	2 g	0,25-1 mm	About 5 months

### Description

Heterologous dry cortico-cancellous collagenic bone mix  
Slightly radiopaque granules

## Characteristics

**Gen-Os®** has a similar structure to human bone (matrix and porous form)<sup>(1)</sup> and presents osteoconductive properties<sup>(2)</sup>.

Thanks to its collagen content, **Gen-Os®** has chemotactic properties and facilitates blood clotting and the subsequent invasion of repairing and regenerative cells<sup>(2)</sup>.

In addition, it is gradually resorbable and provides appropriate support in bone neoformation<sup>(3)</sup> and neo-angiogenesis<sup>(4)</sup>.

**Gen-Os®** is a hydrophilic Dual-Phase xenograft, and it has been also used in association with growth factors<sup>(5)</sup>.

## How to handle Gen-Os®

**Gen-Os®** must always be hydrated and thoroughly mixed with either a few drops of sterile physiological solution or the patient's blood to activate its collagen matrix embedded within the xenogenic bone granules.



1. Figueiredo M et al. J Biomed Mater Res B Appl Biomater, 2010 Feb; 92(2):409-19
2. Nannmark U et al. Clin Implant Dent Relat Res, 2008 Dec;10(4):264-70
3. Cardaropoli D et al. Int J Periodontics Restorative Dent, 2008 Oct; 28(5):469-77
4. Rombouts C et al. Dent Mater J, 2016 Dec 1;35(6):900-907
5. Mijiritsky E et al. Materials, 2017 Sep 8;10(9)



## Porcine origin

Codes	Packaging	Size	Granulometry	Mean observed re-entry time
A1005FS	1 Vial	0,5 g	0,6-1 mm	About 5 months
A1010FS	1 Vial	1 g	0,6-1 mm	About 5 months
A1020FS	1 Vial	2 g	0,6-1 mm	About 5 months
A0210FS	1 Vial	1 g	1-2 mm	About 5 months
AC1005FS	1 Vial	0,5 g	0,6-1 mm	About 5 months
AC1010FS	1 Vial	1 g	0,6-1 mm	About 5 months

## Equine origin

Codes	Packaging	Size	Granulometry	Mean observed re-entry time
A1005FE	1 Vial	0,5 g	0,6-1 mm	About 5 months
A1010FE	1 Vial	1 g	0,6-1 mm	About 5 months
A1020FE	1 Vial	2 g	0,6-1 mm	About 5 months
A0210FE	1 Vial	1 g	1-2 mm	About 5 months

## Description

**Apatos® Mix:** Dry cortico-cancellous collagenic bone mix  
(ref: A1005FS A1010FS A1020FS A0210FS A1005FE A1010FE A1020FE A0210FE)

**Apatos® Cortical:** Dry cortical collagenic bone granules  
(ref: AC1005FS AC1010FS)

Radiopaque granules of mineral hydroxyapatite

## Characteristics

**Apatos®** is a biocompatible and osteoconductive<sup>(1)</sup> collagenic xenograft with characteristics similar to mineralised human bone<sup>(2)</sup>.

The natural microporous consistency of **Apatos®** facilitates the formation of new bone tissue in the bone defect area<sup>(3)</sup>, accelerating the physiological process.

**Apatos® Cortical** is characterised by a long resorption time<sup>(4)</sup>, contributing to the preservation of the grafted volume.

## How to handle Apatos®

**Apatos®** must always be hydrated and thoroughly mixed with a few drops of sterile saline. It can also be mixed with the patient's blood.

The mixture thus obtained should be positioned with a sterile spatula or syringe for biomaterials.



1. Cakir M et al. Int J Oral Maxillofac Implants, 2015 Mar-Apr;30(2):279-85
2. Kolmas J et al. Pharm Biomed Anal, 2012 Mar 5;61:136-41
3. Barone A et al. J Clin Periodontol, 2017 Feb;44(2):204-214
4. Scarano A et al. Clin Implant Dent Relat Res, 2011 Mar;13(1):13-8



## Porcine origin

Codes	Size	Packaging	Mean observed re-entry time
TSV005S	0,5 cc	1 Syringe	About 5 months

### Description

Collagen gel and aqueous solution of a biocompatible copolymer

## Characteristics

**TSV Gel** is a heterologous type I and III collagen gel diluted in aqueous solution containing a biocompatible synthetic copolymer that gives **TSV Gel** thermo-reversible and thermo-gelling properties.

*In vitro* and clinical studies demonstrated that **TSV Gel** components enhance osteoblast activity<sup>(1)</sup> and support new bone formation<sup>(2)</sup>.

At low temperature (+4°C), the gel is relatively flowable and easy to mix with the graft, but it becomes more viscous *in situ* when exposed to body temperature.

## How to handle **TSV Gel**

The purpose of **TSV Gel** is to provide mechanical stability to bone substitutes<sup>(2,3)</sup> and collagen membranes<sup>(4)</sup>.

**TSV Gel** should be refrigerated for at least 20 minutes at +4°C before use, in order to reach the low viscosity phase. It is recommended to mix **TSV Gel** with dry bone granules (e.g. **Gen-Os**<sup>(2)</sup> or **Apatos**<sup>(5)</sup>).



1. Pierfelice TV et al. Int J Mol Sci, 2025 Aug 6;26(15):7621
2. Cinquini C et al. Clin Implant Dent Relat Res, 2025 Aug;27(4):e70085
3. Lopez MA et al. Am J Dent, 2024 Sept;37(SIA):33A-36A
4. Passarelli PC et al. Am J Dent, 2024 Sept;37(SIA):29A-32A



## Equine origin

Codes	Packaging	Size	Mean observed re-entry time
BN0E	1 Blister	10x10x10 mm	About 8 months
BN1E	1 Blister	10x10x20 mm	About 8 months
BN2E	1 Blister	10x20x20 mm	About 8 months
BN8E	1 Blister	35x10x5 mm	About 8 months

### Description

Dry cancellous collagenic bone block

## Characteristics

**Sp-Block** is a xenogenic cancellous block. The bone matrix is not ceramized and the collagen is preserved in its natural structure, thus accelerating physiological resorption and new bone formation<sup>[1-6]</sup>.

**Sp-Block** is able to maintain the original graft volume, which is particularly important in case of large regenerations.

## How to handle Sp-Block

**Sp-Block** must be hydrated for 5/10 minutes before use with a sterile physiological solution.

Afterwards, the block can be adapted to the receiving site which must be accurately decorticated in order to guarantee maximum contact.

The blocks should be always fixed with osteosynthesis microscrews. When used for vertical augmentation with inlay technique, **Sp-Block** should also be fixed with miniplates.

Protection with collagen membranes, like **Evolution**, is recommended. Gaps between the block and bone should be filled with bone substitutes, like **mp3**<sup>[6]</sup>.



1. Scarano A et al. Biomed Res Int, 2016;2016:4086870
2. Sakaguchi R et al. Materials (Basel), 2023 Oct 18;16(20):6742
3. Kaneko N et al. Materials (Basel), 2023 Dec 3;16(23):7490
4. Felice P et al. Int J Periodontics Restorative Dent, 2013 Mar-Apr;33(2):159-66
5. Felice P et al. Int J Periodontics Restorative Dent, 2017 Jul/Aug;37(4):469-480
6. Marconcini S et al. J Oral Maxillofac Surg, 2019 Feb;77(2):289-298



## Porcine origin

Codes	Packaging	Size	Thickness	Mean observed re-entry time
LS10HS	1 Blister	35x35 mm	1 mm	About 6 months
LS23FS	1 Blister	25x35 mm	0,4-0,6 mm	About 5 months
LS25FS	1 Blister	25x25 mm	0,4-0,6 mm	About 5 months
LS24LS	1 Blister	20x40 mm	1 mm	About 6 months
LS03SS	1 Blister	30x30 mm	2-4 mm	About 8 months
LS15LS	1 Blister	35x15 mm	0,6-0,8 mm	About 7 months
LS35LS	1 Blister	35x35 mm	1 mm	About 6 months

## Equine origin

Codes	Packaging	Size	Thickness	Mean observed re-entry time
LS10HE	1 Blister	35x35 mm	1 mm	About 6 months
LS25FE	1 Blister	25x25 mm	0,4-0,6 mm	About 5 months

## Description

Heterologous cortical collagenic bone lamina

**Lamina® Soft:** semi-rigid flexible dried lamina  
(ref: LS25FS LS24LS LS03SS LS25FE LS03SE; Oval LS23FS)

**Lamina® Curved:** semi-rigid flexible dried lamina, curved version  
(ref: LS10HS LS10HE)

**Tecnoss® Lamina® Hard:** rigid dried lamina, flexible after re-hydration  
(ref: LS15LS LS35LS)

## Characteristics

**Lamina®** collagenic bone laminas are made of cortical bone of heterologous origin.

After a process of superficial decalcification, **Lamina® Soft** and **Lamina® Curved** acquires an elastic consistency, nevertheless maintaining the typical compactness of the bone tissue from which it originates; the margins are soft in order not to cause micro-traumas to the surrounding tissue<sup>(1)</sup>.

**Tecnoss® Lamina® Hard** undergoes a process of superficial semi-decalcification therefore increasing its consistency, typical of the cortical bone tissue<sup>(2)</sup>.

## How to handle Lamina®

**Lamina® Soft, Lamina® Curved and Tecnoss® Lamina® Hard** must be fixed with osteosynthesis tools<sup>(1-3)</sup>.

**Lamina® Curved** should always be immobilized<sup>(3)</sup> either with titanium microscrews or sutured directly to the surrounding tissues with a triangular section non-traumatic needle. **Lamina® Curved** should not be hydrated in order to maintain its tenting effect.

**Lamina® Soft** and **Tecnoss® Lamina® Hard** must be hydrated and can be shaped with sterile scissors until the desired size is reached<sup>(1)</sup>, then it must be hydrated for 5/10 minutes in sterile physiological solution.

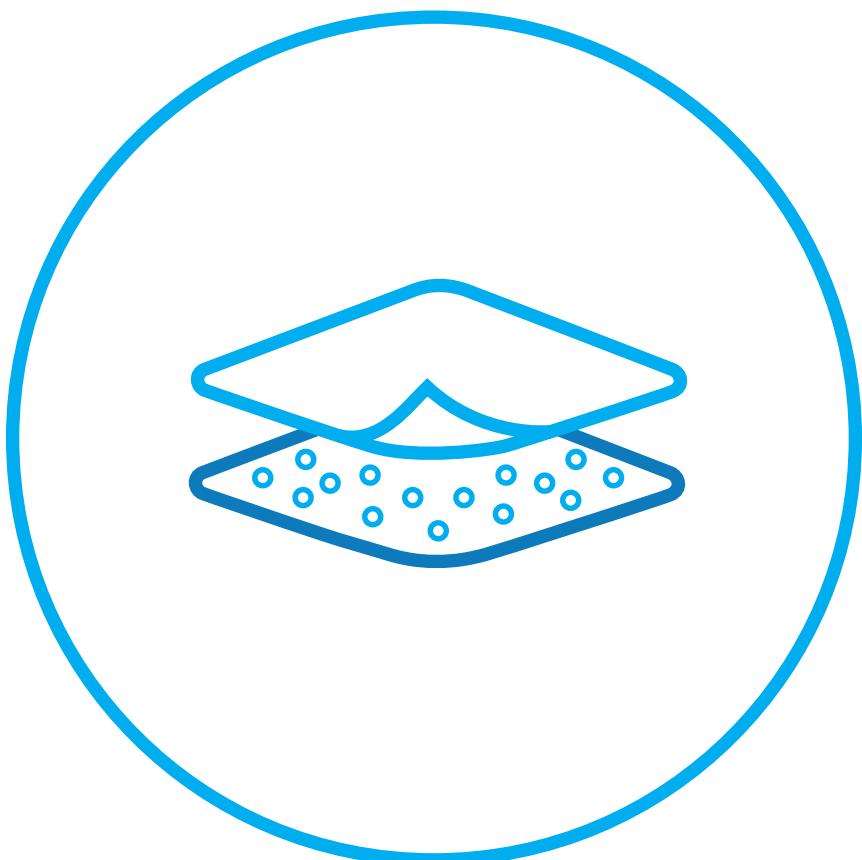
1. Ghallab NA et al. Oral Maxillofac Surg, 2023 Sep;27(3):497-505

2. Happe A et al. J Clin Med. 2023 Nov 9;12(22):7013

3. Rossi R et al. Int J Esthet Dent. 2020;15(4):454-473



# **COLLAGEN MEMBRANES**



# THE ADVANTAGES OF A

## WHY COLLAGEN?

Collagen is the most abundant protein in human bone and one of the main components of the extracellular matrix (ECM), the environment that provides structural and biochemical support to any type of cell<sup>(1)</sup>.

## A NATURAL COLLAGEN BARRIER

The ECM is also a physical barrier that divides one tissue from another. For example, collagen fibres naturally contained within the ECM and in the OsteoBiol® collagen membranes separate osteogenic cells from epithelial cells and fibroblasts<sup>(2)</sup>.

## AVOIDING SOFT TISSUE INGROWTH

Indeed, fibroblasts migration and their subsequent proliferation may jeopardise the work of osteogenic cells. As a consequence, the bone defect is filled by soft tissues and a limited amount of newly formed bone is expected<sup>(3)</sup>; that is the biological rationale behind the use of OsteoBiol® collagen membranes<sup>(4,5)</sup>.

## PROTECTION AGAINST FOOD AND BACTERIA

In addition, the collagen contained within the ECM around gingival cells acts as a barrier against food contamination and bacteria that may proliferate and damage the anatomical structures around teeth or put at risk new bone formation. Scientific studies demonstrate that OsteoBiol® collagen membranes avoid graft contamination, thus allowing successful post-operative healing<sup>(4,5)</sup>.

# COLLAGEN MEMBRANE

The collagen contained within the ECM also acts as a scaffold for the growth of fibroblasts and epithelial cells<sup>(1)</sup>, the two main components of the gingiva.

Being naturally similar to the collagen contained within the ECM, this precious protein retained within the OsteoBiol® collagen membranes allows soft tissues to grow onto *Evolution* and within *Derma* membranes<sup>(3-5)</sup>.

Hence, OsteoBiol® collagen membranes can be used in various surgical procedures to protect and stabilise the graft (*Evolution*) or increase the thickness of the soft tissues (*Derma*).

SUPPORTING SOFT  
TISSUE GROWTH

A NATURAL  
COLLAGEN SCAFFOLD

AN ALTERNATIVE TO  
THE GOLD STANDARD

1. Theocharis AD et al. *Adv Drug Deliv Rev*, 2016 Feb 1;97:4-27
2. Fischer KR et al. *Ann Anat*, 2020 Sep;231:151524
3. Barone A et al. *J Periodontol*, 2014 Jan;85(1):14-23
4. Tallarico M et al. *Eur J Oral Implantol*, 2017;10(2):169-178
5. Fickl S et al. *Clin Oral Implants Res*, 2015 Jul;26(7):799-805

# THE ADVANTAGES OF OUR

## OSTEOBIOL® COLLAGEN MEMBRANES: A UNIQUE PRODUCT LINE

Tecnoss® developed two families of collagen membranes<sup>(1)</sup> (*Evolution* from xenogenic pericardium and peritoneum and *Derma* from porcine dermis) to provide the best handling, the ideal thickness, consistency, stretchability, and optimal regenerative results in adequate re-entry time.

Today it is finally possible to achieve predictable clinical successes in most regenerative protocols<sup>(1)</sup> with the security and support of 25 years of clinical research.



Alternative to CTG for  
soft tissue augmentation



Wide portfolio of membranes



# COLLAGEN MEMBRANES



Graft protection and stabilization are necessary to allow bone and soft tissue regeneration. Various *in vitro* and experimental studies demonstrated that OsteoBiol® collagen membranes have an appropriate resorption time<sup>(2)</sup> to maintain the graft stable and isolated and allow predictable hard and soft tissue healing<sup>(3-8)</sup>.

In some cases, Connective Tissue Graft (CTG), the gold standard for soft tissue regeneration, is necessary to achieve optimal and predictable esthetic outcomes. Because CTG is available in limited quantities and requires to expose patients to multiple surgeries, TecnoSS® designed a collagen membrane to help dentists overcome CTG scarcity: Derma. Dozens of international studies showed that Derma can be used as an alternative to CTG when augmenting soft tissues around teeth and implants and treating mild gingival recessions<sup>(9-12)</sup>.

25 years of clinical experience<sup>(1)</sup> demonstrated that the OsteoBiol® collagen membranes protect the graft, stabilise it<sup>(3-8)</sup> and augment soft tissues<sup>(9-12)</sup> in several surgical procedures.

## PROTECTION AND STABILISATION OF THE GRAFT

## AN ALTERNATIVE TO THE GOLD STANDARD

## A LONG HISTORY OF CLINICAL EXCELLENCE

1. Romasco T et al. *J Funct Biomater*, 2022 Aug 18;13(3):121
2. Vallecillo-Rivas M et al. *Polymers (Basel)*, 2021 Sep 5;13(17):3007
3. Barone A et al. *J Periodontol*, 2014 Jan;85(1):14-23
4. Scarano A et al. *Clin Implant Dent Relat Res*, 2011 Mar;13(1):13-8
5. Barone A et al. *Clin Implant Dent Relat Res*, 2016 Dec;18(6):1103-1112
6. Esposito et al. *Eur J Oral Implantol*, 2015 Autumn;8(3):233-44
7. Piana P et al. *Int J Periodontics Restorative Dent*, 2013 Mar-Apr;33(2):159-66
8. Barone A et al. *Clin Oral Implants Res*, 2015 Jul;26(7):806-13
9. Matoh U et al. *Int J Periodontics Restorative Dent*, 2019;39(3):439-446
10. Fischer K et al. *Clin Implant Dent Relat Res*, 2019;1-8
11. Fischer K et al. *Quintessence Int*, 2014 Nov-Dec;45(10):853-60
12. Minniti A et al. *J Clin Med*, 2024 May 17;13(10):2954



## Porcine origin

Codes	Packaging	Size	Thickness	Mean observed re-entry time
EM03XS	1 Blister	30x30 mm	0,15-0,25 mm	About 2 months
EM02HS	1 Blister	20x20 mm	0,36-0,45 mm	About 4/6 months
EM03HS	1 Blister	30x30 mm	0,36-0,45 mm	About 4/6 months

## Equine origin

Codes	Packaging	Size	Thickness	Mean observed re-entry time
EVO2LLE	1 Blister	20x20 mm	0,26-0,35 mm	About 3/4 months
EVO3LLE	1 Blister	30x30 mm	0,26-0,35 mm	About 3/4 months
EVO2HHE	1 Blister	20x20 mm	0,36-0,45 mm	About 4/6 months
EVO3HHE	1 Blister	30x30 mm	0,36-0,45 mm	About 4/6 months
EVO4LLE	1 Blister	40x40 mm	0,26-0,35 mm	About 3/4 months
EVO6LLE	1 Blister	80x60 mm	0,26-0,35 mm	About 3/4 months

## Description

100% heterologous pericardium or peritoneum tissue

Dried membrane with one smooth side and one micro-rough side

*Evolution X-Fine*: thickness from 0,15 to 0,25 mm  
(ref: EM03XS)

*Evolution Fine*: thickness from 0,26 to 0,35 mm  
(ref: EVO2LLE EVO3LLE EVO4LLE EVO6LLE)

*Evolution Standard*: thickness from 0,36 to 0,45 mm  
(ref: EM02HS EM03HS EVO2HHE EVO2HHE)

## Characteristics

**Evolution** membranes originate from xenogenic pericardium or peritoneum.

The membrane structure, made of dense collagen fibres of high consistency and resistance, makes it a valid choice to protect and stabilise the bone graft in most surgical procedures.

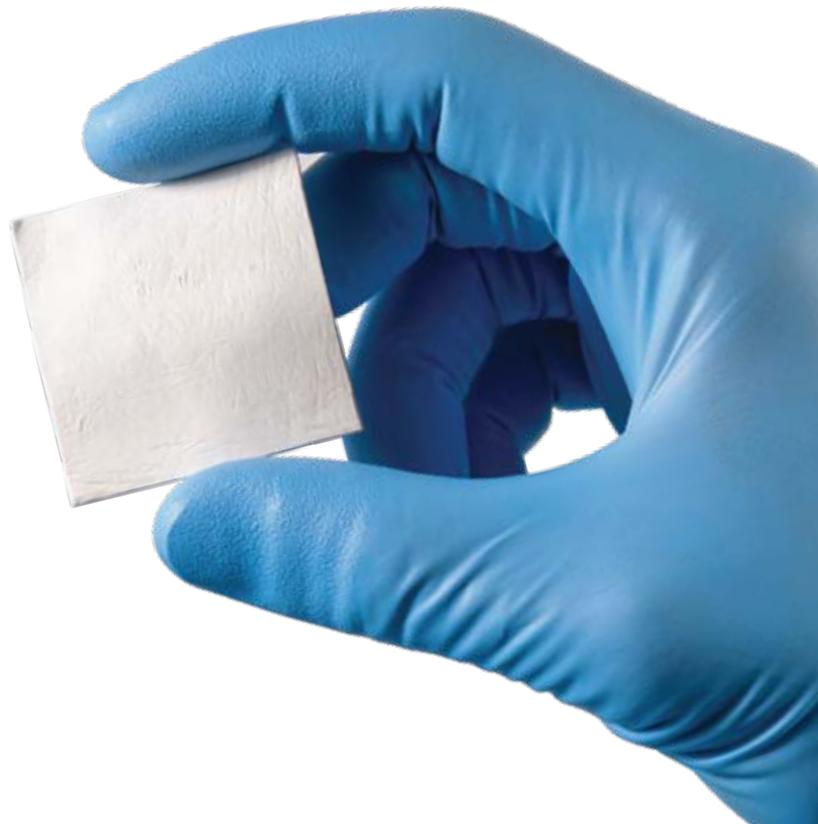
These membranes are gradually resorbable<sup>(1)</sup> and showed great resistance to degradation *in vitro* tests<sup>(2-4)</sup>.

## How to handle Evolution

The membrane can be shaped with sterile scissors until the desired size is reached; unless the grafting site is already bleeding, the membrane should be hydrated with physiological solution.

Once it acquires the desired plasticity, it must be adapted to the grafting site.

In addition, when exposed intentionally or unintentionally, **Evolution** collagen membranes allow second-intention soft tissue healing<sup>(5)</sup>



1. Nannmark U et al. Clin Implant Dent Relat Res, 2008 Dec;10(4):264-70
2. Toledano M et al. Polymers (Basel), 2020 Jun 4;12(6):1290
3. Osorio R et al. Polymers (Basel), 2022 Aug 11;14(16):3274
4. Vallecillo-Rivas M et al. Polymers (Basel), 2021 Sep 5;13(17):3007
5. Barone A et al. J Periodontol, 2014 Jan;85(1):14-23



## Porcine origin

Codes	Packaging	Size	Thickness	Mean observed re-entry time
ED02LS	1 Blister	20x20 mm	0,4-0,6 mm	About 1 month
ED75SS	1 Blister	7x5 mm	2-3 mm	About 5 months
ED15SS	1 Blister	15x5 mm	2-3 mm	About 5 months
ED03SS	1 Blister	30x30 mm	2-3 mm	About 5 months
ED05FS	1 Blister	50x50 mm	0,8-1 mm	About 3 months
ED21FS	1 Blister	12x8 mm (Oval)	0,8-1 mm	About 3 months
ED25FS	1 Blister	25x25 mm	0,8-1 mm	About 3 months
ED05SS	1 Blister	50x50 mm	2-3 mm	About 5 months

## Description

High density collagen membrane from porcine dermis

## Characteristics

**Derma** is a family of collagen membranes that have been used for soft tissue augmentation<sup>(1-8)</sup>.

They are gradually integrated<sup>(1)</sup> with the autologous soft tissues thanks to the presence of natural collagen fibres<sup>(2)</sup>.

Their resistance allows perfect stabilization and prolonged protection of the underlying graft<sup>(3)</sup> in socket regeneration procedures. **Derma** strong consistency guides soft tissue closure and avoids its invagination.

Being an alternative to CTG, **Derma** has been used to augment soft tissues and to improve the quality of keratinized tissues<sup>(4)</sup> around teeth, and implants<sup>(5)</sup> or to treat mild gingival recessions<sup>(6,7)</sup>.

## How to handle Derma

**Derma** can be shaped with scissors and must be hydrated in a sterile physiological solution until the desired shape and consistency are obtained.



1. Fickl S et al. Clin Oral Implants Res, 2015 Jul;26(7):799-805
2. De Marco P et al. Biomed Mater, 2017 Sep 13;12(5):055005
3. Tallarico M et al. Eur J Oral Implantol, 2017;10(2):169-178
4. Fischer K et al. Quintessence Int, 2014 Nov-Dec;45(10):853-60
5. Fischer K et al. Clin Implant Dent Relat Res, 2019;1-8
6. Fickl S et al. Quintessence Int, 2013;44(3):243-6
7. Matoh U et al. Int J Periodontics Restorative Dent, 2019;39(3):439-446



# THE CLINICAL EXPERIENCE



Why choose OsteoBiol® biomaterials



Clinical cases & information



Scientific evidence

A scanning electron micrograph (SEM) showing numerous small, irregularly shaped granules of varying sizes. The granules are light-colored against a darker, textured background, creating a marbled or cellular pattern.

SEM image of GTO® granules  
Source: Politecnico di Torino, Italy

**GTO®**

***The sticky xenograft***

# Why choose GTO®?

---

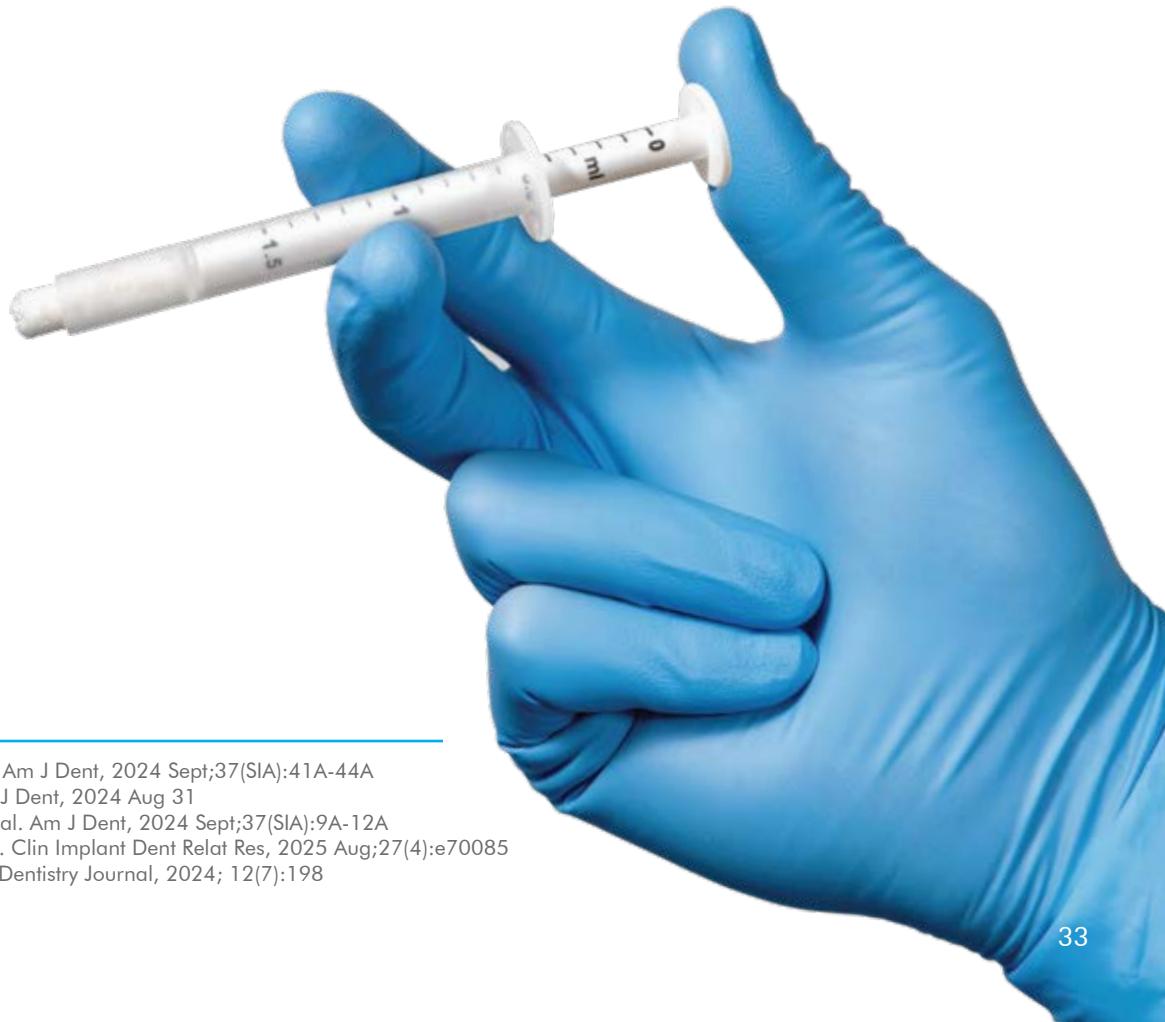
Bone defect conformation varies among patients.

**GTO® is a Dual-Phase sticky xenograft** characterised by high adaptability to the recipient site and high stability once grafted.

**GTO®** allows adequate new bone formation<sup>(1-4)</sup> and socket volume preservation when used for GBR<sup>(5)</sup> in post-extractive sockets with delayed implant placement, even in case of compromised post-extractive sockets.

In combination with **Lamina®**, **GTO® can also be the appropriate choice for the treatment of horizontal defects<sup>(1)</sup> (e.g. knife-edge ridges).**

Thanks to its stickiness and ease of handling, **GTO®** can be used to regenerate bone around peri implant dehiscences<sup>(5)</sup>.



---

1. Lopez MA et al. Am J Dent, 2024 Sept;37(SIA):41A-44A
2. Canullo L et al. J Dent, 2024 Aug 31
3. Passarelli PC et al. Am J Dent, 2024 Sept;37(SIA):9A-12A
4. Cinquini C et al. Clin Implant Dent Relat Res, 2025 Aug;27(4):e70085
5. Menini M et al. Dentistry Journal, 2024; 12(7):198

# Clinical cases & applications

## RECONSTRUCTION OF A SEVERELY COMPROMISED MAXILLA

Sex: female - Age: 19 years old - Author: Dr. Patrick Palacci, private practice, France



1. The entire labial plate of bone is missing. The defect is 16 mm wide, 15 mm height, 5 mm depth.



2. GTO® is placed on top of the ridge with successive layers.



3. Four months later, the newly formed bone is well vascularised and ready to receive an implant treatment.

### HORIZONTAL/VERTICAL AUGMENTATION



## TREATMENT OF A PERIODONTAL DEFECT

Sex: female - Age: 42 years old - Author: Prof. Patrick R. Schmidlin, University of Zurich, Switzerland



1. The pre-operative x-ray shows a deep intrabony defect.



2. GTO® is applied to regenerate the missing bone.



3. The x-ray after six months shows the intrabony defect replaced by an appropriate amount of newly formed bone.

### PERIODONTAL REGENERATION



## RESOLUTION OF A VESTIBULAR DEHISCENCE

Sex: female - Age: 35 years old - Author: Dr. Arturo Gallardo Flores, private practice, Spain



1. The patient reported a vestibular dehiscence in position 46.



2. GTO® is employed to reconstruct the bone vestibularly. A Derma is used as a protective collagen membrane.



3. After four months, an appropriate amount of new bone is regenerated.

### HORIZONTAL AUGMENTATION





## RECONSTRUCTION OF AN ATROPHIC MAXILLA

Sex: male - Age: 77 years old - Author: Prof. Tiziano Testori, University of Michigan, USA



1. The pre-operative intraoperative view shows an edentulous area in the posterior maxilla.



2. Implants are placed simultaneously with GTO® after sinus floor elevation.



3. The clinical view reveals that implants are stable after five years.

### LATERAL ACCESS SINUS LIFT



## TREATMENT OF A COMPROMISED POST-EXTRACTION SOCKET

Sex: male - Age: 47 years old - Author: Prof. Antonio Barone, University of Pisa, Italy



1. Tooth extraction leads to a deep post-extractive socket lacking the buccal bone.

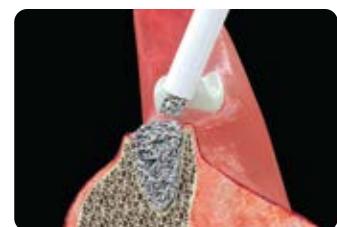


2. The defect is grafted with GTO® and the missing buccal bone is reconstructed with Lamina® Soft.

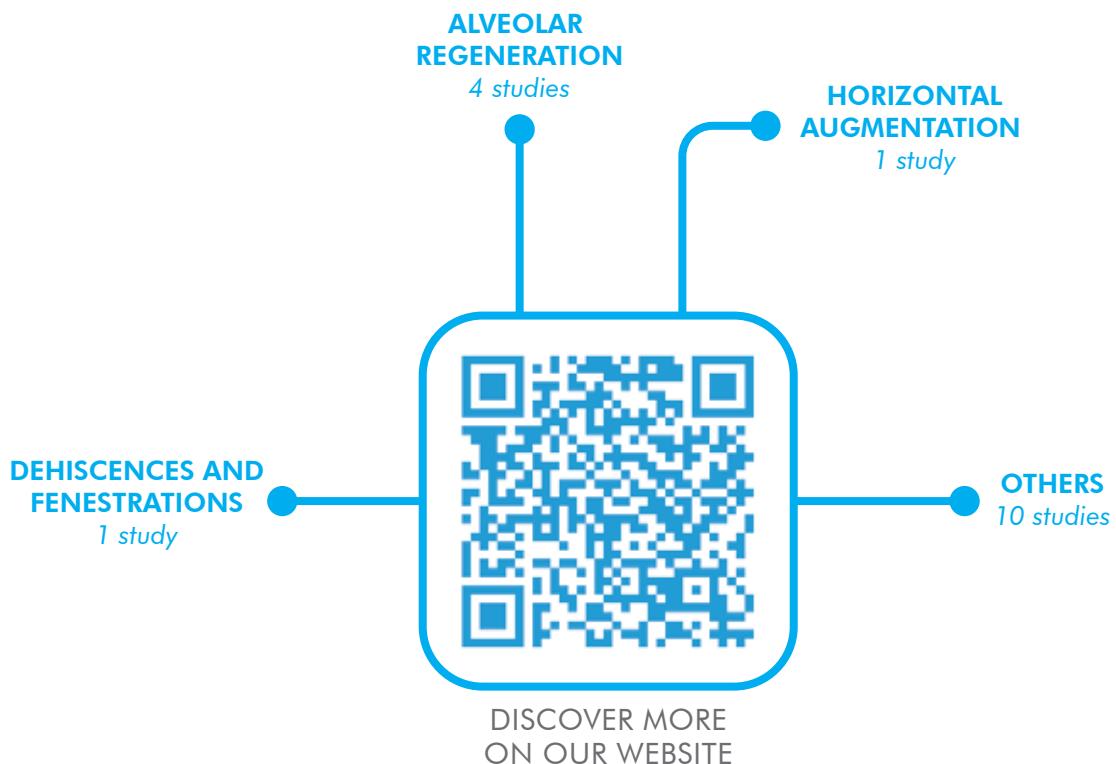


3. After six months, the site is ready to receive the implant.

### ALVEOLAR REGENERATION



# GTO®'s scientific evidence



## SELECTED SCIENTIFIC BIBLIOGRAPHY

### ALVEOLAR REGENERATION

Canullo L, Savadori P, Triestino A, Pesce P, Sora V, Caponio VCA, Mangano F, Menini M.  
**Investigation of the extent of post-extraction bone contraction and remodeling after 4 months. A prospective pilot study.**  
J Dent. 2024 Nov;150:105337

Passarelli PC, Lopez MA, Netti A, Felicetti A, Wychowański P, Garcia-Godoy F, Chiapasco M, D'Addona A.  
**Alveolar regeneration of the post-extraction site with cortical deficiency using the Lamina Socket Sealing Technique: a retrospective study with clinical, radiographic and histomorphometric analysis. Part 1/2.**  
Am J Dent. 2024 Sept;37(SIA):4A-8A

Passarelli PC, Lopez MA, Netti A, Felicetti A, Wychowański P, Garcia-Godoy F, Chiapasco M, D'Addona A.  
**Alveolar regeneration of the post-extraction site with cortical deficiency using the Lamina Socket Sealing Technique: retrospective study with histomorphometric analysis of regenerated bone and evaluation of soft tissue changes. Part 2/2.**  
Am J Dent. 2024 Sept;37(SIA):9A-12A

Cinquini C, D'amico E, Iezzi G, Priami M, Santarelli S, Barone A.  
**Cortico-cancellous collagenic porcine bone for alveolar ridge preservation: a cohort comparative study.**  
Clin Implant Dent Relat Res. 2025 Aug;27(4):e70085

### DEHISCENCES AND FENESTRATIONS

Menini M, Canullo L, Iacono R, Triestino A, Caponio VCA, Savadori P, Pesce P, Pedetta A, Guerra F.  
**Effect of different graft material consistencies in the treatment of minimal bone dehiscence: a retrospective pilot study.**  
Dent J (Basel). 2024 Jun 27;12(7):198

### EXPERIMENTAL STUDIES

Taniguchi Z, Esposito M, Xavier SP, Silva ER, Botticelli D, Buti J, Baba S.  
**On the use of a sticky bone substitute in the presence of a ruptured Schneider membrane during sinus lift procedures - An experimental within-rabbit study.**  
Int J Oral Maxillofac Implants. 2024 Oct 4;0(0):1-28

### HORIZONTAL AUGMENTATION

Lopez MA, Passarelli PC, Netti A, Marianelli F, Wychowański P, Garcia-Godoy F, D'Addona A.  
**A new technique of alveolar bone augmentation: "Pack into Bone". A retrospective case-series study.**  
Am J Dent. 2024 Sept;37(SIA):41A-44A

### IN VITRO STUDIES

Di Tinco R, Consolo U, Pisciotta A, Orlandi G, Bertani G, Nasi M, Bertacchini J, Carnevale G.  
**Characterization of dental pulp stem cells response to bone substitutes biomaterials in dentistry.**  
Polymers (Basel). 2022 May 30;14(11):2223

Jeanneau C, Catherine JH, Giraud T, Lan R, About I.  
**The added value of a collagenated thermosensitive bone substitute as a scaffold for bone regeneration.**  
Materials (Basel). 2024 Jan 27;17(3):625

Pierfelice TV, Cinquini C, Petrini M, D'Amico E, D'Arcangelo C, Barone A, Iezzi G.  
**Evaluation of collagenic porcine bone blended with a collagen gel for bone regeneration: an in vitro study.**  
Int J Mol Sci. 2025 Aug 6;26(15):7621

Pelaez-Cruz P, López Jornet P, Pons-Fuster E.  
**Collagen formulation in xenogeneic bone substitutes influences cellular responses in periodontal regeneration: an in vitro study.**  
Biomimetics. 2025; 10(9):608

#### REVIEWS

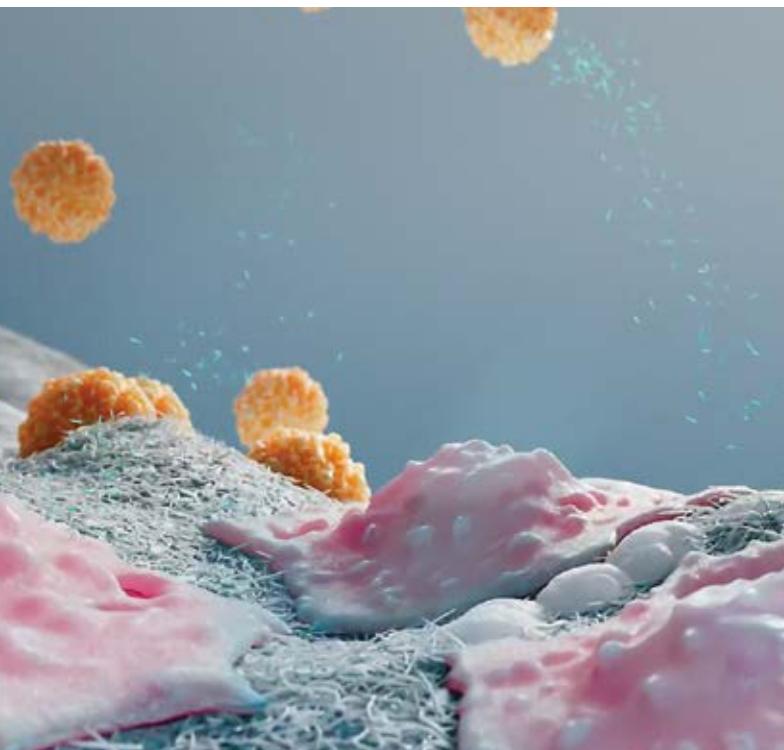
Saleh MHA, Sabri H, Di Pietro N, Comuzzi L, Geurs NC, Bou Semaan L, Piattelli A.

**Clinical indications and outcomes of sinus floor augmentation with bone substitutes: an evidence-based review.**

Clin Implant Dent Relat Res. 2024 Oct 17

# OsteoBiol® GTO® by Tecnoss

## THE STICKY XENOGRAFT



**OUTSTANDING STABILITY DUE TO IDEAL VISCOSITY**

**EASILY ADAPTABLE TO THE RECIPIENT SITE**

**READY-TO-USE IN STERILE SYRINGE**

**INNOVATIVE DUAL-PHASE XENOGRAFTS**

- TSV GEL INSIDE FOR **OPTIMAL STICKINESS**
- **DIRECTLY INJECTABLE** INTO BONE DEFECT
- **FASTER AND SAFER** SURGICAL HANDLING
- CORTICO-CANCELLOUS **COLLAGENIC** MATRIX

A scanning electron micrograph (SEM) showing numerous small, irregularly shaped granules of mp3®. The granules vary in size and density, creating a complex, textured surface. The colors range from light blue to white, with darker blue highlights on the more prominent peaks and valleys of the granule clusters.

SEM image of mp3® granules  
Source: Politecnico di Torino, Italy

mp3®

*The injectable  
Dual-Phase xenograft*

# Why choose mp3®?

Although autogenous bone (AB) is still the gold standard for bone regeneration, its harvesting is an invasive and time-consuming process.

**mp3® is a ready-to-use Dual-Phase biomaterial** that can be directly injected into sinus cavities, showing comparable results to autogenous bone chips<sup>(1,2)</sup>.

In addition, **mp3®** can be easily grafted into post-extractive sockets to preserve the ridge volume<sup>(3)</sup> and allow a correct second-stage implant placement. **mp3®** can be used in combination with **Evolution** membranes<sup>(1-3)</sup>.

Thanks to its similarity to natural human bone<sup>(1,4)</sup>, **mp3® allows adequate graft volume preservation<sup>(3)</sup>**, a healthy new bony tissue formation<sup>(1)</sup> and ultimately, **a successful implant rehabilitation over time<sup>(1,5-7)</sup>**.



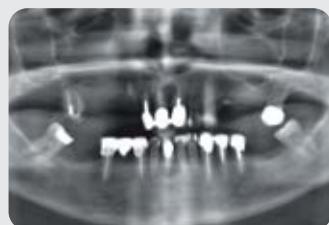
---

1. Correia et al. Materials (Basel), 2021 Jun 21;14(12):3439
2. Correia et al. Dentistry Journal, 2024; 12(2):33
3. Barone A et al. Clin Implant Dent Relat Res. 2017 Aug;19(4):750-759
4. Figueiredo M et al. J Biomed Mater Res B Appl Biomater, 2010 Feb; 92(2):409-19
5. Pistilli R et al. J Dent, 2022 Jun;121:104137
6. Buonocunto N et al. Clin Implant Dent Relat Res, 2023 Dec;25(6):1044-1055
7. Cinquini C et al. Clin Implant Dent Relat Res, 2024 Dec;26(6):1162-1171

# Clinical cases & applications

## LATERAL ACCESS SINUS LIFT AND HORIZONTAL AUGMENTATION

Sex: female - Age: 50 years old - Author: Prof. Tiziano Testori, University of Michigan, USA



1. Based on the pre-operative radiograph, all teeth are considered hopeless.



2. Upon sinus membrane elevation, mp3® is injected into the antrostomy.



3. Six months later, implants are placed in a high-quality bone tissue.

### LATERAL ACCESS SINUS LIFT/ HORIZONTAL AUGMENTATION



## TREATMENT OF A COMPROMISED ALVEOLAR SOCKET

Sex: female - Age: 54 years old - Author: Prof. Antonio Barone, University of Pisa, Italy



1. The pre-operative clinical view shows a fractured tooth.



2. mp3® is grafted into the post-extractive socket and protected with a Derma collagen membrane.



3. After seven years, the clinical view shows no sign of peri-implantitis or mucositis.

### ALVEOLAR REGENERATION



## TREATMENT OF A HORIZONTAL DEFECT

Sex: Female - Age: 43 years old - Author: Prof. Michael Weinländer, University of Graz, Austria



1. Three implants are positioned in a narrow ridge.



2. mp3® and Lamina® Soft are used to reconstruct the defect.



3. The clinical view shows newly formed bone after eight months.

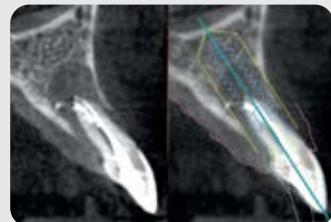
### HORIZONTAL AUGMENTATION





## THE MULTI-LAYER TECHNIQUE

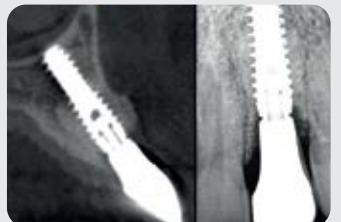
Sex: female - Age: 38 years old - Author: Dr Iñaki Gamborena, private practice, Spain



1. The CBCT shows that both the buccal and the palatal bone are missing.



2. *mp3®* is packed around the implant. The buccal and palatal bone are reconstructed with *Lamina® Soft*.



3. The four-year follow-up radiograph shows an implant surrounded by vital bone.

### MULTI-LAYER TECHNIQUE



## THE MODIFIED PAPILLA PRESERVATION TECHNIQUE

Sex: male - Age: 61 years old - Author: Dr. Gerd Körner, private practice, Germany



1. The initial x-ray shows a deep periodontal defect (9 mm depth).



2. The defect is filled with *mp3®* and covered with a tailored *Evolution* membrane.

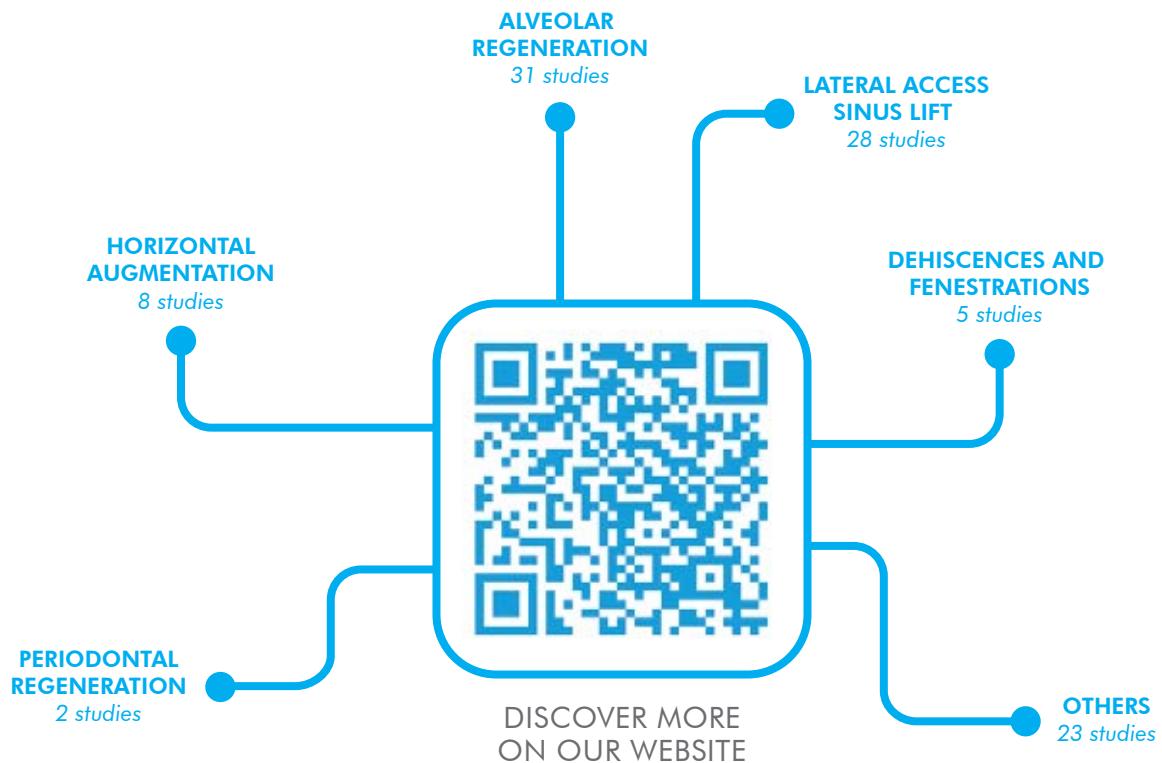


3. After four months, the intrabony defect is resolved.

### PERIODONTAL REGENERATION



# mp3®'s scientific evidence



## SELECTED SCIENTIFIC BIBLIOGRAPHY

### ALVEOLAR REGENERATION

Barone A, Orlando B, Cingano L, Marconcini S, Derchi G, Covani U. **A randomized clinical trial to evaluate and compare implants placed in augmented versus non-augmented extraction sockets: 3-year results.** J Periodontol. 2012 Jul;83(7):836-46

Barone A, Ricci M, Tonelli P, Santini S, Covani U. **Tissue changes of extraction sockets in humans: a comparison of spontaneous healing vs. ridge preservation with secondary soft tissue healing.** Clin Oral Implants Res. 2013 Nov;24(11):1231-7

Barone A, Borgia V, Covani U, Ricci M, Piattelli A, Iezzi G. **Flap versus flapless procedure for ridge preservation in alveolar extraction sockets: a histological evaluation in a randomized clinical trial.** Clin Oral Implants Res. 2015 Jul;26(7):806-13

Barone A, Ricci M, Romanos GE, Tonelli P, Alfonsi F, Covani U. **Buccal bone deficiency in fresh extraction sockets: a prospective single cohort study.** Clin Oral Implants Res. 2015 Jul;26(7):823-30

Thalmair T, Fickl S, Schneider D, Hinze M, Wachtel H. **Dimensional alterations of extraction sites after different alveolar ridge preservation techniques - A volumetric study.** J Clin Periodontol. 2013 Jul;40(7):721-7

Barone A, Toti P, Menchini-Fabris GB, Derchi G, Marconcini S, Covani U.

**Extra oral digital scanning and imaging superimposition for volume analysis of bone remodeling after tooth extraction with and without 2 types of particulate porcine mineral insertion: a randomized controlled trial.**

Clin Implant Dent Relat Res. 2017 Aug;19(4):750-759

Corbella S, Taschieri S, Francetti L, Weinstein R, Del Fabbro M. **Histomorphometric results after postextraction socket healing with different biomaterials: a systematic review of the literature and meta-analysis.** Int J Oral Maxillofac Implants. 2017 Sep/Oct;32(5):1001-1017

Barone A, Toti P, Piattelli A, Iezzi G, Derchi G, Covani U. **Extraction socket healing in humans after ridge preservation techniques: comparison between flapless and flapped procedures in a randomized clinical trial.** J Periodontol. 2014 Jan;85(1):14-23

Marconcini S, Gianniaro E, Derchi G, Alfonsi F, Covani U, Barone A.

**Clinical outcomes of implants placed in ridge-preserved versus nonpreserved sites: a 4-year randomized clinical trial.**

Clin Implant Dent Relat Res. 2018 Dec;20(6):906-914

Schuh PL, Wachtel H, Beuer F, Goker F, Del Fabbro M, Francetti L, Testori T.  
**Multi-Layer Technique (MLT) with porcine collagenated cortical bone lamina for bone regeneration procedures and immediate post-extraction implantation in the esthetic area: a retrospective case series with a mean follow-up of 5 years.**  
Materials (Basel). 2021 Sep 9;14(18):5180

Körner G, Bäumer-König A, Barz-Popp F, Tripodakis A.  
**Esthetic implant solutions in the periodontally compromised anterior region. Expanding the indications for immediate implant placement with the Socket Rebuilding Technique (SRT).**  
Int J Esthet Dent. 2022 Nov 25;17(4):408-422

Buonocunto N, Cinquini C, Mijiritsky E, Tagger-Green N, Porreca A, Di Nicola M, Iezzi G, Barone A.  
**Effect of alveolar ridge preservation on peri-implant mucositis and peri-implantitis prevalence: a multicenter, cross-sectional study.**  
Clin Implant Dent Relat Res. 2023 Jun 19

Cinquini C, Izzetti R, Porreca A, Iezzi G, Nisi M, Barone A.  
**Alveolar ridge preservation and its impact on marginal bone level changes around dental implants: a retrospective, cohort comparative study.**  
Clin Implant Dent Relat Res. 2024 Aug 19

#### DEHISCENCES AND FENESTRATIONS

Slote C, Lindfors N, Nannmark U.  
**Surgical reconstruction of peri-implant bone defects with prehydrated and collagenated porcine bone and collagen barriers: case presentations.**  
Clin Implant Dent Relat Res. 2013 Oct;15(5):714-23

#### EXPERIMENTAL STUDIES

Nannmark U, Sennerby L.  
**The bone tissue responses to prehydrated and collagenated cortico-cancellous porcine bone grafts: a study in rabbit maxillary defects.**  
Clin Implant Dent Relat Res. 2008 Dec;10(4):264-70

Falacho RI, Palma PJ, Marques JA, Figueiredo MH, Caramelo F, Dias I, Viegas C, Guerra F.  
**Collagenated porcine heterologous bone grafts: histomorphometric evaluation of bone formation using different physical forms in a rabbit cancellous bone model.**  
Molecules. 2021 Mar 2;26(5):1339

Taniguchi Z, Esposito M, Xavier SP, Silva ER, Botticelli D, Buti J, Baba S.  
**On the use of a sticky bone substitute in the presence of a ruptured Schneider membrane during sinus lift procedures - An experimental within-rabbit study.**  
Int J Oral Maxillofac Implants. 2024 Oct 4;0(0):1-28

#### HORIZONTAL AND VERTICAL AUGMENTATION

Wachtel H, Fickl S, Hinze M, Bolz W, Thalmair T.  
**The Bone Lamina Technique: a novel approach for lateral ridge augmentation - A case series.**  
Int J Periodontics Restorative Dent. 2013 Jul-Aug;33(4):491-7

Marconcini S, Covani U, Giammarinaro E, Velasco-Ortega E, De Santis D, Alfonsi F, Barone A.  
**Clinical success of dental implants placed in posterior mandible augmented with interpositional block graft: 3-year results from a prospective cohort clinical study.**  
J Oral Maxillofac Surg. 2019 Feb;77(2):289-298

#### LATERAL ACCESS SINUS LIFT

Correia F, Pozza DH, Gouveia S, Felino AC, Faria-Almeida R.  
**Advantages of porcine xenograft over autograft in sinus lift: a randomised clinical trial.**  
Materials (Basel). 2021 Jun 21;14(12):3439

Pistilli R, Canullo L, Pesce P, Pistilli V, Caponio C, Sbricoli L.  
**Guided implant surgery and sinus lift in severely resorbed maxillae: a retrospective clinical study with up to 10 years of follow-up.**  
J Dent. 2022 Apr 21;121:104137

Correia F, Gouveia SA, Pozza DH, Felino AC, Faria-Almeida R.  
**A randomized clinical trial comparing implants placed in two different biomaterials used for maxillary sinus augmentation.**  
Materials (Basel). 2023; 16(3):1220

Correia F, Gouveia S, Felino AC, Faria-Almeida R, Pozza DH.  
**Maxillary sinus augmentation with xenogenic collagen-retained heterologous cortico-cancellous bone: a 3-year follow-up randomized controlled trial.**  
Dent J 2024, 12, 33

#### REVIEWS

Saleh MHA, Sabri H, Di Pietro N, Comuzzi L, Geurs NC, Bou Semaan L, Piatelli A.  
**Clinical indications and outcomes of sinus floor augmentation with bone substitutes: an evidence-based review.**  
Clin Implant Dent Relat Res. 2024 Oct 17

# Gel 40

***The injectable  
Dual-Phase bone gel***

# Why choose **Gel 40**?

---

The perforation of the Schneiderian membrane is one of the major complications that may occur during transcrestal sinus floor elevation<sup>(1)</sup>.

Various studies reported that **Gel 40** simplifies the transcrestal sinus floor elevation<sup>(2-6)</sup> with immediate implant positioning: the collagen gel contained in **Gel 40** gently detaches the sinus membrane making the surgery easier, faster<sup>(4)</sup>, and predictable<sup>(4,5)</sup>.

Finally, in case of accidental perforation of the Schneiderian membrane, **Gel 40** bone granules accidentally dispersed into the sinus are easily extruded without major complications<sup>(6)</sup>.



---

1. Chen MH et al. J Prosthodont, 2018 Jun;27(5):394-401
2. Barone A et al. Int J Periodontics Restorative Dent, 2008 Jun;28(3):283-9
3. Santagata M et al. J Oral Implantol, 2010;36(6):485-9
4. Lombardi T et al. Int J Implant Dent, 2022 Jul 22;8(1):32
5. Lopez MA et al. Am J Dent, 2024 Sept;37(SIA):25A-28A
6. Bernardello F et al. Sinusitis, 2021; 5(2):132-140

# Clinical cases & applications

DISCOVER MORE



ON OUR WEBSITE

## CRESTAL SINUS LIFT WITH IMMEDIATE IMPLANT PLACEMENT

Sex: male - Age: 59 years old - Authors: Prof. Claudio Stacchi, University of Trieste, Italy and Dr. Fabio Bernardello, private practice, Italy



1. An implant was inserted to replace tooth in position 16 about ten years before. The implant subject to this case was placed in 17.



2. Gel 40 is injected into the antrostomy.



3. Seven months after the surgery, Gel 40 remodeled into new bone above the new implant apex.

## CRESTAL ACCESS SINUS LIFT



## TREATMENT A PERIODONTAL DEFECT

Sex: female - Age: 49 years old - Author: Dr. Tom Kobe, private practice, Slovenia



1. The patient shows a deep periodontal defect.



2. Gel 40 is injected into the defect to regenerate bone around the tooth root.



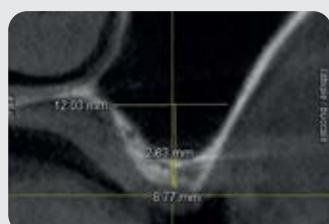
3. At one-year follow-up, the bone has been reconstructed.

## PERIODONTAL REGENERATION



## TRANSCRESTAL SINUS FLOOR ELEVATION

Sex: female - Age: 55 years old - Author: Dr. Fabio Bernardello, private practice, Italy



1. Based on the sinus width, the patient is considered eligible for a transcrestal sinus floor elevation.



2. Gel 40 is used to detach the Schneiderian membrane. Immediate implant insertion prevents the biomaterial extrusion.



3. Gel 40 has been remodeled into new bone above the new implant.

## CRESTAL ACCESS SINUS LIFT



# Gel 40's scientific evidence



## SELECTED SCIENTIFIC BIBLIOGRAPHY

### CRESTAL ACCESS SINUS LIFT

Barone A, Cornelini R, Ciaglia R, Covani U.

**Implant placement in fresh extraction sockets and simultaneous osteotome sinus floor elevation: a case series.**

Int J Periodontics Restorative Dent. 2008 Jun;28(3):283-9

Santagata M, Guariniello L, Rauso R, Tartaro G.

**Immediate loading of dental implant after sinus floor elevation with osteotome technique: a clinical report and preliminary radiographic results.**

J Oral Implantol. 2010;36(6):485-9

Comuzzi L, Tumedei M, Piattelli A, Tartaglia G, Del Fabbro M. **Radiographic analysis of graft dimensional changes in transcrestal maxillary sinus augmentation: a retrospective study.**

Materials (Basel). 2022 Apr 19;15(9):2964

Lombardi T, Lamazza L, Bernardello F, Ziętek G, Stacchi C, Troiano G. **Clinical and radiographic outcomes following transcrestal maxillary sinus floor elevation with injectable xenogenous bone substitute in gel form: a prospective multicenter study.**

Int J Implant Dent. 2022 Jul 22;8(1):32

Lopez MA, Passarelli PC, Netti A, Felicetti A, Dell'Aquila F, Wychowański P, D'Addona A.

**Transcrestal maxillary sinus floor elevation with injectable xenogeneic bone substitute in gel form: a clinical, radiological and histological analysis.**

Am J Dent. 2024 Sept;37(SIA):25A-28A

### EXPERIMENTAL STUDIES

Nannmark U, Azarmehr I.

**Short communication: collagenated cortico-cancellous porcine bone grafts. A study in rabbit maxillary defects.**

Clin Implant Dent Relat Res. 2010 Jun 1;12(2):161-3

### PERIODONTAL REGENERATION

Cardaropoli D, Cardaropoli G.

**Healing of gingival recessions using a collagen membrane with a hemineralized xenograft: a randomized controlled clinical trial.**

Int J Periodontics Restorative Dent. 2009 Feb;29(1):59-67

Kobe T, Povšič K, Gašperšič R.

**Prehydrated collagenated cortico-cancellous heterologous bone gel and papillae tunneling for isolated intrabony defects: 12-month noninferiority trial.**

Clin Exp Dent Res. 2024 Feb;10(1):e853

# **Putty**

***The injectable  
Dual-Phase bone paste***

# Why choose **Putty**?

---

Peri-implant and small self-containing bone defects can be effectively grafted with a viscous biomaterial with small-diameter granules.

The “soft” consistency, the small-diameter bone granules of **Putty** and the practical ready-to-use syringe packaging make it a valid choice for small self-containing peri-implant lesions<sup>(1, 2)</sup> and small defects with a self-containing cavity.

Thanks to its collagen gel content, **Putty** gently detaches the sinus membrane, thus being a valid option for transcrestal sinus floor elevation<sup>(3)</sup> with immediate implant placement.



---

1. Barone A et al. Eur J Implant Prosthodontics, 2006, 2:99-106

2. Cassetta M et al. Int J Periodontics Restorative Dent, 2012 Oct;32(5):581-9

3. Saglanmak A et al. J Clin Med, 2024 Apr 11;13(8):2225

# Clinical cases & applications

DISCOVER MORE



ON OUR WEBSITE

## TREATMENT OF A PERI-IMPLANT LESION

Sex: male - Age: 59 years old - Author: Dr. Emilio Rodríguez Fernández, private practice, Spain



1. The defect and the implant surface are cleaned and decontaminated.



2. The peri-implant lesion is filled with Putty.



3. Three months later, the peri-implant lesion is resolved.

### PERI-IMPLANT LESION



## TRANSCRESTAL SINUS FLOOR ELEVATION

Sex: female - Age: 51 years old - Author: Dr. Luca Comuzzi, private practice, Italy



1. The initial x-ray shows a bone defect suitable for a transcrestal sinus elevation.



2. The Schneiderian membrane is gently elevated with Putty.



3. The twelve-year follow-up radiograph shows an implant surrounded by vital bone.

### CRESTAL ACCESS SINUS LIFT



## INFERIOR ALVEOLAR NERVE (IAN) TRANSPOSITION AND HORIZONTAL AUGMENTATION

Sex: female - Age: 43 years old - Authors: Prof. Raquel Zita and Dr. Hugo Moreira, private practices, Portugal



1. The initial panoramic x-ray shows the mental foramen near the bone ridge.



2. Lamina® is placed after grafting a mixture of Putty, mp3® and Plasma Rich Fibrin (PRF).

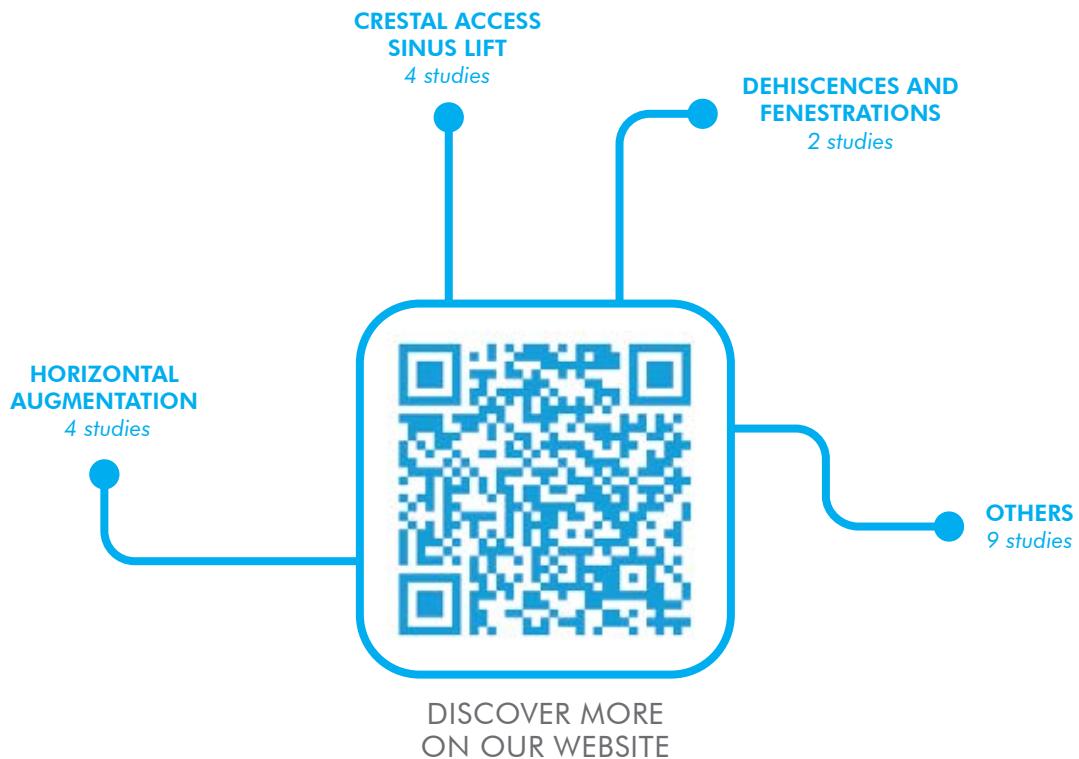


3. The panoramic x-ray shows that all implants are placed and stable.

### HORIZONTAL AUGMENTATION



# Putty's scientific evidence



## SELECTED SCIENTIFIC BIBLIOGRAPHY

### CRESTAL ACCESS SINUS LIFT

Lopez MA, Lico S, Casale M, Ormanier Z, Carinci F. **The use of various biomaterials in computer-guided crestal sinus lift procedures. A report on two case studies with volume comparison.** Oral Implantol (Rome). 2016 Nov 13;9(2):89-97

Comuzzi L, Tumedei M, Piattelli A, Tartaglia G, Del Fabbro M. **Radiographic analysis of graft dimensional changes in transcrestal maxillary sinus augmentation: a retrospective study.** Materials (Basel). 2022 Apr 19;15(9):2964

Saglanmak A, Cinar IC, Zboun M, Arisan V, Mijiritsky E. **Maxillary sinus floor elevation and simultaneous implant installation via osseodensification drills: a retrospective analysis of bone gain in 72 patients followed for 6 months.** J Clin Med. 2024 Apr 11;13(8):2225

### DEHISCENCES AND FENESTRATIONS

Cassetta M, Ricci L, Iezzi G, Dell'Aquila D, Piattelli A, Perrotti V. **Resonance frequency analysis of implants inserted with a simultaneous grafting procedure: a 5-year follow-up study in man.** Int J Periodontics Restorative Dent. 2012 Oct;32(5):581-9

### EXPERIMENTAL STUDIES

Nannmark U, Azarmehr I. **Short communication: collagenated cortico-cancellous porcine bone grafts. A study in rabbit maxillary defects.** Clin Implant Dent Relat Res. 2010 Jun 1;12(2):161-3

### HORIZONTAL AUGMENTATION

Santagata M, Guariniello L, Tartaro G. **A modified edentulous ridge expansion technique for immediate placement of implants: a case report.** J Oral Implantol. 2011 Mar;37 Spec No:114-9

### REVIEWS

Saleh MHA, Sabri H, Di Pietro N, Comuzzi L, Geurs NC, Bou Semaan L, Piattelli A. **Clinical indications and outcomes of sinus floor augmentation with bone substitutes: an evidence-based review.** Clin Implant Dent Relat Res. 2024 Oct 17

A scanning electron micrograph (SEM) showing numerous irregular, white, porous granules of Gen-Os®. The granules have a complex, sponge-like structure with many small pores and ridges, giving them a textured appearance. They are scattered across a light blue background.

SEM image of **Gen-Os®** granules  
Source: Politecnico di Torino, Italy

# Gen-Os®

***The advantages of a  
Dual-Phase xenograft***

# Why choose Gen-Os®?

Collagen is a precious protein for bone regeneration.

**Gen-Os®** is the first Dual-Phase xenogenic bone substitute developed by Tecnoss® physically and chemically similar to autogenous bone<sup>(1,2)</sup>.

**Gen-Os®** can be used alone or mixed with AB chips whenever necessary<sup>(3-5)</sup> for pre-implant and peri-implant regeneration procedures.

In addition, due to its granulometry range, **Gen-Os®** has been successfully used for the treatment of deep and narrow periodontal defects<sup>(6-10)</sup>.

Finally, more than 25 years of clinical experience and about 100 scientific studies showed that **Gen-Os®** is a valid universal bone filler that allows predictable results in various surgical procedures<sup>(3-11)</sup>.



---

1. University of Duisburg-Essen, Germany
2. Figueiredo M et al. J Biomed Mater Res B Appl Biomater, 2010 Feb; 92(2):409-19
3. Pellegrino G al al. Int J Oral Implantol (Berl), 2024 May 27;17(2):175-185
4. Felice P et al. Int J Oral Implantol (Berl), 2024 Sep 16;17(3):285-296
5. Elaskary A et al. BMC Oral Health, 2024 Oct 7;24(1):1190
6. Cardaropoli D et al. Int J Periodontics Restorative Dent, 2009 Feb;29(1):59-67
7. Esposito M et al. Eur J Oral Implantol, 2015 Autumn;8(3):233-44
8. Aslan S et al. J Clin Periodontol, 2017 Sep;44(9):926-932 – n200
9. Aslan S et al. Int J Periodontics Restorative Dent, 2017 Mar/Apr;37(2):227-233
10. Kobe T et al. Clin Exp Dent Res, 2024 Feb;10(1):e853
11. Romasco T et al. J Funct Biomater, 2022 Aug 18;13(3):121

# Clinical cases & applications

## TREATMENT OF A PERIODONTAL DEFECT

Sex: male - Age: 40 years old - Author: Dr. Tom Kobe: private practice, Slovenia



1. A vertical incision (EPPT) at the adjacent tooth is done to expose the defect.



2. The defect is filled with Gen-Os®.



3. The one-year follow-up shows the resolution of the bone defect.

### PERIODONTAL REGENERATION



## MANAGEMENT OF A DEHISCENCE

Sex: female - Age: 45 years old - Authors: Prof. Fernando Guerra and Dr. Rui Falacho, University of Coimbra, Portugal



1. At the time of implant placement, a dehiscence is reported.



2. Gen-Os® is grafted to regenerate new bone.



3. The three-year follow up shows the implant surrounded by vital bone.

### DEHISCENCES AND FENESTRATIONS



## TRANSCRESTAL SINUS FLOOR ELEVATION

Sex: female - Age: 66 years old - Author: Prof. Claudio Stacchi, University of Trieste, Italy



1. The pre-operative x-ray shows 2,5 mm residual bone height and the presence of an Underwood septum.



2. Sinus is grafted by condensing Gen-Os® with osteotomes, until a minimum height of 10 mm was obtained.



3. The clinical view shows newly formed bone after eight months.

### CRESTAL ACCESS SINUS LIFT





## GBR AND DELAYED IMPLANT PLACEMENT IN THE ESTHETIC ZONE

Sex: female - Age: 68 years old - Author: Dr. Irfan Abas, private practice, The Netherlands



1. After retracting the flap, the inflammatory tissue is removed.



2. Alveoli are reconstructed with sticky bone (Gen-Os® + Plasma Rich Fibrin).



3. A sufficient thickness of tissues for a natural emergence profile is achieved.

### ALVEOLAR REGENERATION



## AUTOTRANSPLANTED TOOTH WITH BONE FENESTRATION\*

Sex: male - Age: 16 years old - Authors: Dr. Francesco Giachi Carù, private practice, Italy and Prof. Tiziano Testori, University of Michigan, USA



1. The superior cuspid is unrecoverable with traditional orthodontic treatment.



2. The cuspid is transplanted and the bone is reconstructed with sticky bone (Gen-Os® + Plasma Rich Fibrin).



3. The five-year follow-up clinical view shows no sign of complications in the area of the transplanted cuspid.

### ALVEOLAR REGENERATION



## TREATMENT OF A WIDE HORIZONTAL DEFECT

Sex: female - Age: 45 years old - Author: Dr. Pavel Yaroshevich, private practice, Russia



1. In the posterior maxilla, a full-thickness, slightly buccal flap is elevated in the keratinized gingiva.



2. Gen-Os® is placed buccally and linguinally on the defect to reconstruct the bone horizontally.



3. An increase in bone width is observed six months after augmentation.

### HORIZONTAL AUGMENTATION



# Gen-Os®'s scientific evidence



## SELECTED SCIENTIFIC BIBLIOGRAPHY

### ALVEOLAR REGENERATION

Cardaropoli D, Cardaropoli G.

**Preservation of the postextraction alveolar ridge: a clinical and histologic study.**

Int J Periodontics Restorative Dent. 2008 Oct;28(5):469-77

Festa VM, Addabbo F, Laino L, Femiano F, Rullo R.

**Porcine-derived xenograft combined with a soft cortical membrane versus extraction alone for implant site development: a clinical study in humans.**

Clin Implant Dent Relat Res. 2013 Oct;15(5):707-13

Esposito M, Barausse C, Bonifazi L, Piattelli M, Pistilli R, Ferri A, Felice P.

**Wide diameter immediate post-extractive implants versus socket preservation and delayed placement of normal-diameter implants in the molar region: 5-year post-loading outcome of a randomised controlled trial.**

Clinical Trials In Dentistry, 2020;02(4):21-37

Corbella S, Taschieri S, Francetti L, Weinstein R, Del Fabbro M. **Histomorphometric results after postextraction socket healing with different biomaterials: a systematic review of the literature and meta-analysis.**

Int J Oral Maxillofac Implants. 2017 September/October;32(5):1001-1017

Testori T, Simion M, Carù FG, Corbella S, Taschieri S, Perrotti G, Cocconi R, Calatrava J, Decker A, Wang HL.

**Immature tooth autotransplantation with buccal bone fenestration treated with GBR and immature tooth autotransplantation in regenerated bone: proof of principles with medium - (55 months) to long-term (252 months) follow-ups.**

Int J Periodontics Restorative Dent. 2024 Dec 20;0(0):1-22

### DEHISCENCES AND FENESTRATIONS

Cassetta M, Ricci L, Iezzi G, Dell'Aquila D, Piattelli A, Perrotti V. **Resonance frequency analysis of implants inserted with a simultaneous grafting procedure: a 5-year follow-up study in man.**

Int J Periodontics Restorative Dent. 2012 Oct;32(5):581-9

### EXPERIMENTAL STUDIES

Nannmark U, Sennerby L.

**The bone tissue responses to prehydrated and collagenated cortico-cancellous porcine bone grafts: a study in rabbit maxillary defects.**

Clin Implant Dent Relat Res. 2008 Dec;10(4):264-70

Figueiredo A, Coimbra P, Cabrita A, Guerra F, Figueiredo M. **Comparison of a xenogeneic and an alloplastic material used in dental implants in terms of physico-chemical characteristics and in vivo inflammatory response.**

Mater Sci Eng C Mater Biol Appl. 2013 Aug 1;33(6):3506-13

Fischer KR, Stavropoulos A, Calvo-Guirado JL, Schneider D, Fickl S. **Influence of local administration of pamidronate on extraction socket healing - A histomorphometric proof-of-principle pre-clinical in vivo evaluation.**

Clin Oral Implants Res. 2015 Oct;26(10):1135-42

Omori Y, Ricardo Silva E, Botticelli D, Apaza Alccayhuaman KA, Lang NP, Xavier SP.  
**Reposition of the bone plate over the antrostomy in maxillary sinus augmentation: a histomorphometric study in rabbits.**  
Clin Oral Implants Res. 2018 Aug;29(8):821-834

Falacho RI, Palma PJ, Marques JA, Figueiredo MH, Caramelo F, Dias I, Viegas C, Guerra F.  
**Collagenated porcine heterologous bone grafts: histomorphometric evaluation of bone formation using different physical forms in a rabbit cancellous bone model.**  
Molecules. 2021 Mar 2;26(5):1339

Miyauchi Y, Izutani T, Teranishi Y, Iida T, Nakajima Y, Xavier SP, Baba S.  
**Healing patterns of non-collagenated bovine and collagenated porcine xenografts used for sinus floor elevation: a histological study in rabbits.**  
J Funct Biomater. 2022 Dec 5;13(4):276

Nakajima Y, Botticelli D, De Rossi EF, Ferreira Balan V, Pires Godoy E, Ricardo Silva E, Xavier SP.  
**Schneiderian membrane collateral damage caused by collagenated and non-collagenated xenografts: a histological study in rabbits.**  
Dent J (Basel). 2023 Jan 26;11(2):31

#### **HORIZONTAL/VERTICAL AND VERTICAL AUGMENTATION**

Rossi R, Ghezzi C, Tomecek M.  
**Cortical lamina: a new device for the treatment of moderate and severe tridimensional bone and soft tissue defects.**  
Int J Esthet Dent. 2020;15(4):454-473

Villa G, Rizzacasa A, Bessa L, Spina P, Barootchi S, Tavelli L, Wang HL.  
**Shell Technique with a xenogeneic cortical bone Lamina and particulate bone graft for horizontal ridge augmentation: a case series.**  
Int J Periodontics Restorative Dent. 2023 July-Aug;43(4):435-441

Pellegrino G, Vignudelli E, Barausse C, Bonifazi L, Renzi T, Tayeb S, Felice P.  
**Accuracy of semi-occlusive CAD/CAM titanium mesh using the reverse guided bone regeneration digital protocol: a preliminary clinical study.**  
Int J Oral Implantol (Berl). 2024 May 27;17(2):175-185

Felice P, Pistilli R, Pellegrino G, Bonifazi L, Tayeb S, Simion M, Barausse C.  
**A randomised controlled trial comparing the effectiveness of guided bone regeneration with polytetrafluoroethylene titanium-reinforced membranes, CAD/CAM semi-occlusive titanium meshes and CAD/CAM occlusive titanium foils in partially atrophic arches.**  
Int J Oral Implantol (Berl). 2024 Sep 16;17(3):285-296

#### **IN VITRO STUDIES**

Figueiredo M, Henriques J, Martins G, Guerra F, Judas F, Figueiredo H.  
**Physicochemical characterization of biomaterials commonly used in dentistry as bone substitutes-comparison with human bone.**  
J Biomed Mater Res B Appl Biomater. 2010 Feb;92(2):409-419

Rombouts C, Jeanneau C, Camilleri J, Laurent P, About I.  
**Characterization and angiogenic potential of xenogeneic bone grafting materials: role of periodontal ligament cells.**  
Dent Mater J. 2016 Dec 1;35(6):900-907

Mijiritsky E, Ferroni L, Gardin C, Bressan E, Zanette G, Piattelli A, Zavan B.  
**Porcine bone scaffolds adsorb growth factors secreted by MSCs and improve bone tissue repair.**  
Materials (Basel). 2017 Sep 8;10(9):1054

Jeanneau C, Le Fournis C, About I.  
**Xenogeneic bone filling materials modulate mesenchymal stem cell recruitment: role of the Complement C5a.**  
Clin Oral Investig. 2020 Jul;24(7):2321-2329

#### **LATERAL ACCESS SINUS LIFT**

Barone A, Crespi R, Aldini NN, Fini M, Giardino R, Covani U.  
**Maxillary sinus augmentation: histologic and histomorphometric analysis.**  
Int J Oral Maxillofac Implants. 2005 Jul-Aug;20(4):519-25

Barone A, Santini S, Sbordone L, Crespi R, Covani U.  
**A clinical study of the outcomes and complications associated with maxillary sinus augmentation.**  
Int J Oral Maxillofac Implants. 2006 Jan-Feb;21(1):81-5

Cassetta M, Perrotti V, Calasso S, Piattelli A, Sinjari B, Iezzi G.  
**Bone formation in sinus augmentation procedures using autologous bone, porcine bone, and a 50: 50 mixture: a human clinical and histological evaluation at 2 months.**  
Clin Oral Implants Res. 2015 Oct;26(10):1180-4

Tanaka K, Iezzi G, Piattelli A, Ferri M, Mesa NF, Apaza Alccayhuaman KA, Botticelli D.  
**Sinus floor elevation and antrostomy healing: a histomorphometric clinical study in humans.**  
Implant Dent. 2019 Dec;28(6):537-542

Imai H, Iezzi G, Piattelli A, Ferri M, Apaza Alccayhuaman KA, Botticelli D.  
**Influence of the dimensions of the antrostomy on osseointegration of mini-implants placed in the grafted region after sinus floor elevation: a randomized clinical trial.**  
Int J Oral Maxillofac Implants. 2020 May/Jun;35(3):591-598

Hirota A, Iezzi G, Piattelli A, Ferri M, Tanaka K, Apaza Alccayhuaman KA, Botticelli D.  
**Influence of the position of the antrostomy in sinus floor elevation on the healing of mini-implants: a randomized clinical trial.**  
Oral Maxillofac Surg. 2020 Sep;24(3):299-308

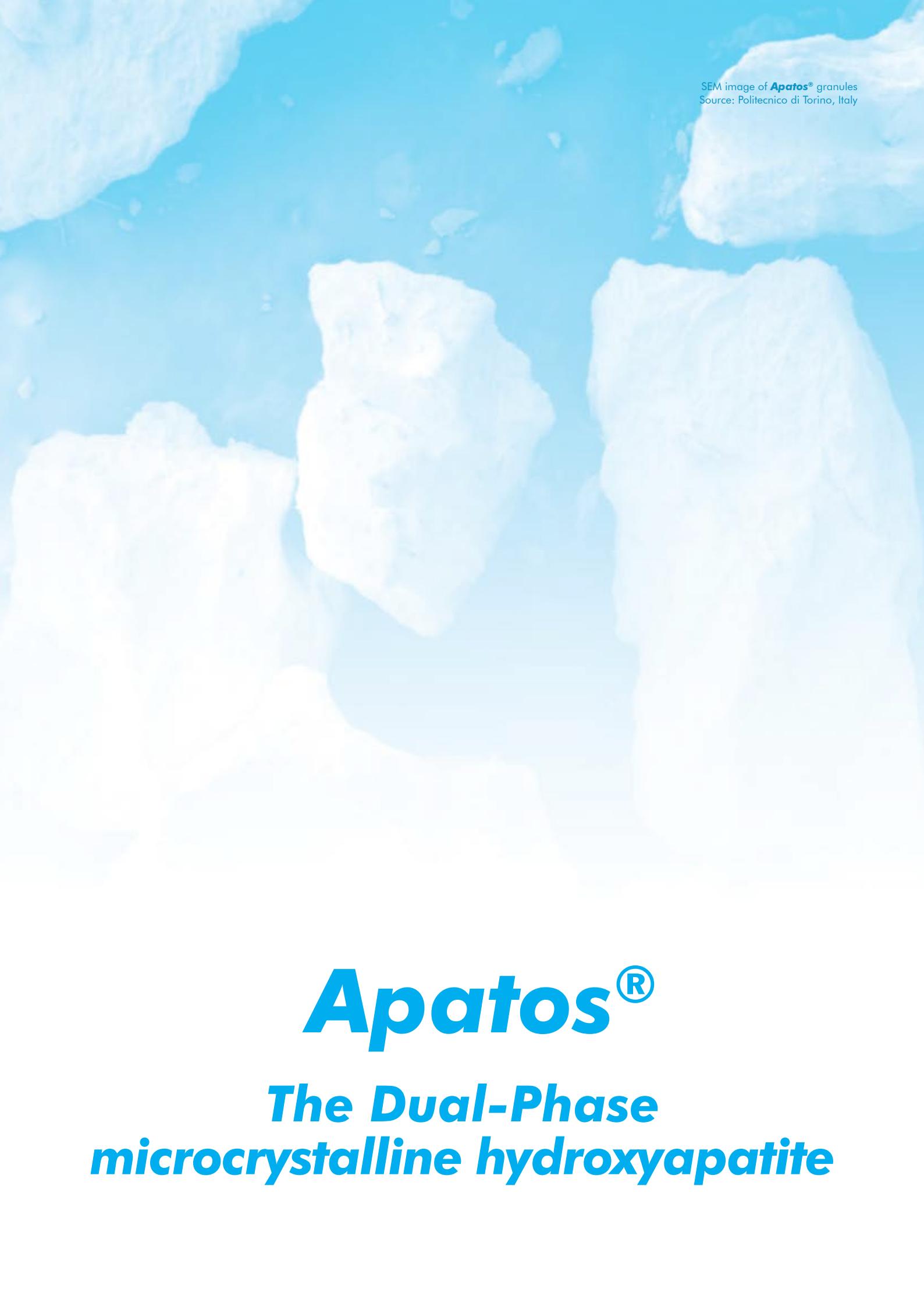
Testori T, Scaini R, Friedland B, Saibene AM, Felisati G, Craig JR, Deflorian M, Zuffetti F, Del Fabbro M, Wang HL.  
**Maxillary sinus opacification after surgery in asymptomatic patients: transient swelling of the sinus mucosa or graft dispersion into the maxillary sinus. A radiographic report of three cases after a follow-up period of at least 5 years.**  
Int J Oral Implantol (Berl). 2024 May 27;17(2):189-198

#### **PERIODONTAL REGENERATION**

Esposito M, Grusovin MG, Lambert F, Matos S, Pietruska M, Rossi R, Salhi L, Buti J.  
**The effectiveness of a resorbable bone substitute with a resorbable membrane in the treatment of periodontal infrabony defect - A multicenter randomised controlled trial.**  
Eur J Oral Implantol. 2015 Autumn;8(3):233-44

Aslan S, Buduneli N, Cortellini P.  
**Entire Papilla Preservation Technique: a novel surgical approach for regenerative treatment of deep and wide intrabony defects.**  
Int J Periodontics Restorative Dent. 2017 Mar/Apr;37(2):227-233

Kobe T, Povšič K, Gašperšič R.  
**Prehydrated collagenated cortico-cancellous heterologous bone gel and papillae tunneling for isolated intrabony defects: 12-month noninferiority trial.**  
Clin Exp Dent Res. 2024 Feb;10(1):e853

A scanning electron micrograph (SEM) showing numerous irregular, white, granular particles of Apatos®. These particles vary in size and density, with some appearing more porous or crystalline than others. They are set against a dark, textured background.

SEM image of **Apatos®** granules  
Source: Politecnico di Torino, Italy

# **Apatos®**

## ***The Dual-Phase microcrystalline hydroxyapatite***

# Why choose **Apatos®**?

---

Because of its capacity to preserve the volume of bone defects<sup>(1,2)</sup> and its medium-size granulometry, **Apatos®** is the proper ally for dentists' daily surgical practice.

Hence, **Apatos®** has been used as a valid alternative to autologous bone to regenerate and preserve the volume of bony lesions, treat two-wall defects<sup>(3)</sup> and molar or premolar extraction sockets<sup>(2)</sup>.

When needed, **Apatos®** can be protected with **Evolution** membrane<sup>(4)</sup> or stabilised with **Lamina®<sup>(5)</sup>**.



---

1. Menini M et al. Dentistry Journal, 2024; 12(7):198
2. Barone A et al. Clin Implant Dent Relat Res, 2017 Aug;19(4):750-759
3. Barone A et al. Clin Implant Dent Relat Res, 2015 Dec;17(6):1114-26
4. Marconcini S et al. Clin Implant Dent Relat Res, 2018 Dec;20(6):906-914
5. Köttgen C et al. Int J Esthet Dent, 2024 May 10;19(2):152-169

# Clinical cases & applications

DISCOVER MORE



ON OUR WEBSITE

## RECONSTRUCTION OF AN ATROPHIC MANDIBLE

Sex: female - Age: 45 years old - Author: Dr. Christopher Köttgen, private practice, Germany



1. The missing bony wall is reconstructed with Tecnooss® Lamina® Hard.



2. The defect is filled with a mix of 60% autogenous bone and 40% Apatos®.



3. The occlusal view of the regenerated area shows two stable implants surrounded by healthy soft tissue.

### VERTICAL AUGMENTATION



## TREATMENT OF A DEHISCENCE

Sex: female - Age: 42 years old - Author: Dr. Pablo Neira Vazquez, private practice, Spain



1. At the time of implant placement, a dehiscence is reported.



2. Apatos® is grafted to regenerate new bone.



3. The six-month follow-up clinical view shows an uneventful healing.

### DEHISCENCES AND FENESTRATIONS



## MANAGEMENT OF A POST-EXTRACTIVE SOCKET

Sex: male - Age: 25 years old - Authors: Dr. Antonio Acevedo and Dr. Antonio Maté De Haro, private practice, Spain



1. The fractured tooth is removed.



2. The post-extractive socket is filled with Apatos®. An Evolution membrane is used to cover and stabilise the graft.

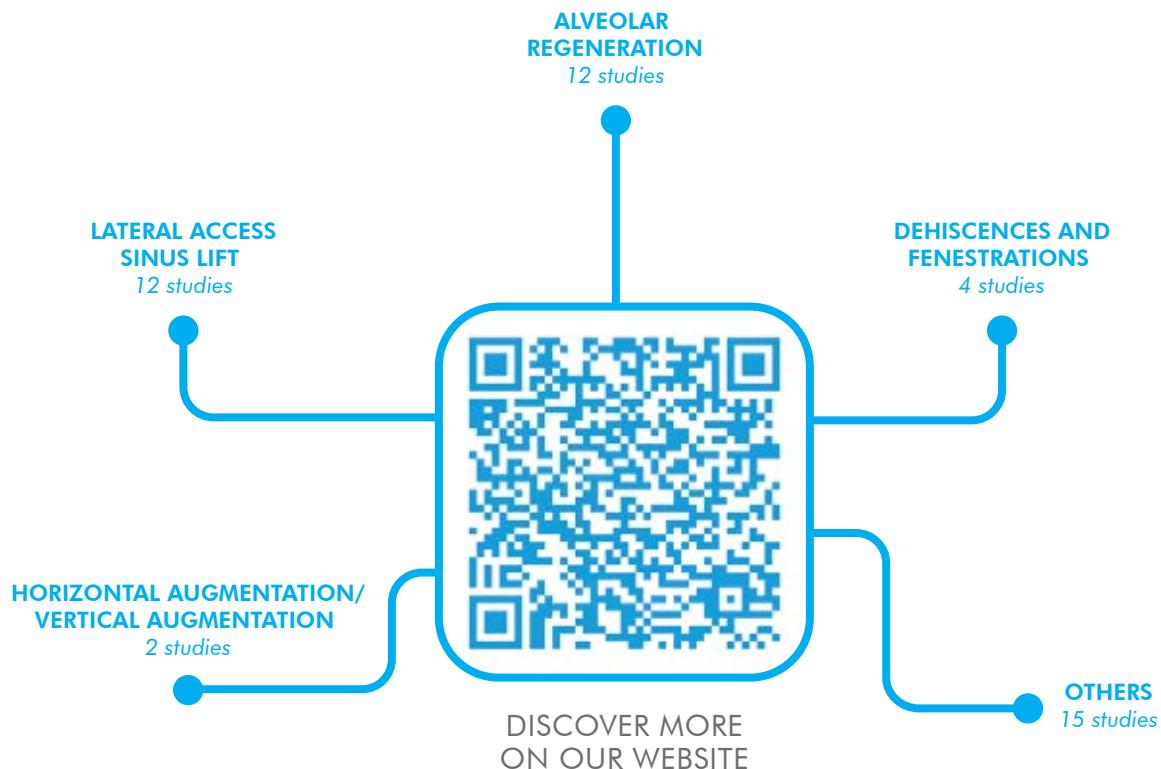


3. An implant is placed in an appropriate amount of newly formed bone four months after surgery.

### ALVEOLAR REGENERATION



# Apatos®'s scientific evidence



## SELECTED SCIENTIFIC BIBLIOGRAPHY

### ALVEOLAR REGENERATION

Barone A, Toti P, Menchini-Fabris GB, Derchi G, Marconcini S, Covani U.  
**Extra oral digital scanning and imaging superimposition for volume analysis of bone remodeling after tooth extraction with and without 2 types of particulate porcine mineral insertion: a randomized controlled trial.**  
Clin Implant Dent Relat Res. 2017 Aug;19(4):750-759

Corbella S, Taschieri S, Francetti L, Weinstein R, Del Fabbro M.  
**Histomorphometric results after postextraction socket healing with different biomaterials: a systematic review of the literature and meta-analysis.**  
Int J Oral Maxillofac Implants. 2017 September/October; 32(5):1001-1017

Marconcini S, Giammarinaro E, Derchi G, Alfonsi F, Covani U, Barone A.  
**Clinical outcomes of implants placed in ridge-preserved versus nonpreserved sites: a 4-year randomized clinical trial.**  
Clin Implant Dent Relat Res. 2018 Dec;20(6):906-914

Köttgen C, Köttgen I, Plaster U.  
**Immediate implant placement in a dialysis patient. A case report.**  
Int J Esther Dent. 2024 May 10;19(2):152-169

### DEHISCENCES AND FENESTRATIONS

Barone A, Toti P, Quaranta A, Derchi G, Covani U.  
**The clinical outcomes of immediate versus delayed restoration procedures on immediate implants: a comparative cohort study for single-tooth replacement.**  
Clin Implant Dent Relat Res. 2015 Dec;17(6):1114-26

Covani U, Canullo L, Toti P, Alfonsi F, Barone A.  
**Tissue stability of implants placed in fresh extraction sockets: a 5-year prospective single-cohort study.**  
J Periodontol. 2014 Sep;85(9):e323-32

### LATERAL ACCESS SINUS LIFT

Scarano A, Piattelli A, Assenza B, Quaranta A, Perrotti V, Piattelli M, Iezzi G.  
**Porcine bone used in sinus augmentation procedures: a 5-year retrospective clinical evaluation.**  
J Oral Maxillofac Surg. 2010 Aug;68(8):1869-73

Scarano A, Piattelli A, Perrotti V, Manzon L, Iezzi G.  
**Maxillary sinus augmentation in humans using cortical porcine bone: a histological and histomorphometrical evaluation after 4 and 6 months.**  
Clin Implant Dent Relat Res. 2011 Mar;13(1):13-8

Iezzi G, Degidi M, Piattelli A, Mangano C, Scarano A, Shibli JA, Perrotti V.  
**Comparative histological results of different biomaterials used in sinus augmentation procedures: a human study at 6 months.**  
Clin Oral Implants Res. 2012 Dec;23(12):1369-76

# **Sp-Block**

## ***The Dual-Phase cancellous block***

# Why choose **Sp-Block**?

---

Vertical augmentation usually requires an extensive amount of autogenous bone, which increases surgical time and may cause more post-operative pain to the patient and increase the risk of morbidity.

In case of vertical augmentation with the inlay technique, the time to harvest an adequate amount of autogenous bone can be spared by using **Sp-Block**.

This Dual-Phase cancellous block can be used as an alternative to autogenous bone blocks when a vertical gain in the posterior mandible is required<sup>(1-4)</sup> and the amount of residual crestal bone allows the inlay technique protocol.



---

1. Felice P et al. Int J Periodontics Restorative Dent, 2013 Mar-Apr;33(2):159-66
2. Barone A et al. J Craniomaxillofac Surg, 2017 Sep;45(9):1438-1447
3. Felice P et al. Int J Periodontics Restorative Dent, 2017 Jul/Aug;37(4):469-480
4. Marconcini S et al. J Oral Maxillofac Surg, 2019 Feb;77(2):289-298

# Clinical cases & applications

DISCOVER MORE



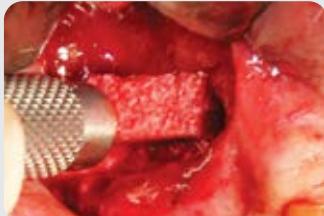
ON OUR WEBSITE

## VERTICAL AUGMENTATION WITH OSTEOTOMY

Sex: male - Age: 77 years old - Authors: Dr Rustam Shamsutdinov and Dr. Ilgam Urazbakhin, private practice, Russia.



1. A horizontal osteotomy is performed in the upper jaw ridge.



2. The bone is augmented with Sp-Block.



3. Two implants are inserted in the augmented ridge.

### VERTICAL AUGMENTATION



## THE INLAY TECHNIQUE

Sex: female - Age: 70 years old - Author: Prof. Pietro Felice, University of Bologna, Italy



1. The pre-operative clinical image shows an atrophic posterior right mandible.



2. The pristine bone is elevated. Sp-Block is placed in the obtained space.



3. After one year, implants are stable and soft tissue is healthy.

### VERTICAL AUGMENTATION

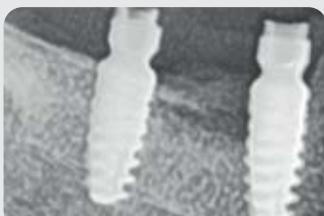


## THE INLAY TECHNIQUE

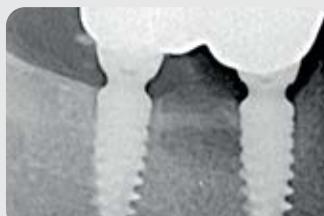
Sex: female - Age: 70 years old - Authors: Prof. Pietro Felice and Dr. Carlo Barausse, University of Bologna, Italy



1. The OPG shows the partially edentulous posterior mandible.



2. An implant is placed in an appropriate amount of newly formed bone four months after the surgery.



3. After five years, the implants are osseointegrated and stable.

### VERTICAL AUGMENTATION



# Sp-Block's scientific evidence



## SELECTED SCIENTIFIC BIBLIOGRAPHY

### EXPERIMENTAL STUDIES

Sakaguchi R, Xavier SP, Morinaga K, Botticelli D, Silva ER, Nakajima Y, Baba S.  
**Histological comparison of collagenated cancellous equine bone blocks used as inlay or onlay for lateral bone augmentation in rabbits.**  
Materials (Basel). 2023 Oct 18;16(20):6742

Kaneko N, Xavier SP, Morinaga K, Botticelli D, Silva ER, Nakajima Y, Baba S.  
**Implants placed with a ring technique using inlay and onlay block xenografts in the mandible of rabbits.**  
Materials (Basel). 2023 Dec 3;16(23):7490

### VERTICAL AUGMENTATION

Scarano A, Carinci F, Assenza B, Piattelli M, Murmura G, Piattelli A.  
**Vertical ridge augmentation of atrophic posterior mandible using an Inlay Technique with a xenograft without miniscrews and miniplates: case series.**  
Clin Oral Implants Res. 2011 Oct;22(10):1125-1130

Felice P, Piana L, Checchi L, Pistilli R, Pellegrino G.  
**Vertical ridge augmentation of the atrophic posterior mandible with a 2-stage Inlay Technique: a case report.**  
Implant Dent. 2012 Jun;21(3):190-5

Felice P, Piana L, Checchi L, Corvino V, Nannmark U, Piattelli M.  
**Vertical ridge augmentation of an atrophic posterior mandible with an Inlay Technique and cancellous equine bone block: a case report.**  
Int J Periodontics Restorative Dent. 2013 Mar-Apr;33(2):159-66

Barone A, Toti P, Menchini-Fabris GB, Felice P, Marchionni S, Covani U.  
**Early volumetric changes after vertical augmentation of the atrophic posterior mandible with interpositional block graft versus onlay bone graft: a retrospective radiological study.**  
J Craniomaxillofac Surg. 2017 Sep;45(9):1438-1447

Felice P, Barausse C, Barone A, Zucchelli G, Piattelli M, Pistilli R, Ippolito DR, Simion M.  
**Interpositional augmentation technique in the treatment of posterior mandibular atrophies: a retrospective study comparing 129 autogenous and heterologous bone blocks with 2 to 7 years follow-up.**  
Int J Periodontics Restorative Dent. 2017 Jul/Aug;37(4):469-480

Marconcini S, Covani U, Giammarinaro E, Velasco-Ortega E, De Santis D, Alfonsi F, Barone A.  
**Clinical success of dental implants placed in posterior mandible augmented with interpositional block graft: 3-year results from a prospective cohort clinical study.**  
J Oral Maxillofac Surg. 2019 Feb;77(2):289-298

Scarano A, Khater AGA, Falisi G, Gehrke SA, Tari SR.  
**Atrophic anterior mandible treated with sandwich osteotomy without mini-screws and mini-plates: a case report with 7 years of follow-up.**  
Clin Case Rep. 2025 Apr 9;13(4):e70377

# Lamina®

***The Dual-Phase  
cortical bone Lamina***

# Why choose **Lamina®**?

---

Autogenous bone (AB) cortical plates harvesting is a complex and invasive procedure and may lead to pain at the donor site, increasing the surgical time and cost. In addition, it may happen that such plates do not fully adapt to the anatomy of the defect to treat.

Under the name **Lamina®** oral surgeons can find a family of heterologous collagenic bone lamina that can be used as an alternative to AB cortical plates in various surgical procedures.

**Lamina® Soft** is a viable option to reconstruct the missing buccal bone in the esthetic area, allowing an immediate implant placement<sup>(1-4)</sup>, high PES<sup>(3,4)</sup> and alveolar contour preservation<sup>(2,4)</sup>.

**Lamina® Soft** and **Lamina® Curved** are also a well-documented choices to treat horizontal defects or knife-edge ridges in combination with Dual-Phase biomaterials<sup>(5)</sup>.

Finally, **TecnoSS® Lamina® Hard** is a valid alternative to AB cortical bone plates for vertical augmentation with the Modified Shell Technique<sup>(6)</sup>.



---

1. Schuh P et al. Materials, 2021, 14, 5180
2. Elaskary A et al. Clin Implant Dent Relat Res, 2022 Oct;24(5):559
3. Ghallab NA et al. Oral Maxillofac Surg, 2023 Sep;27(3):497-505
4. Elaskary A et al. Int J Oral Maxillofac Implants, 2023 May, Jun;38(3):468-478
5. Rossi R et al. Int J Esthet Dent, 2020;15(4):454-473
6. Happe A et al. J Clin Med, 2023 Nov 9;12(22):7013

# Clinical cases & applications

## COMPLEX RECONSTRUCTION OF AN ATROPHIC MAXILLA

Sex: female - Age: 53 years old - Author: Dr. Iñaki Gamborena, private practice, Spain



1. The clinical image shows the virtual representation of the full contour wax-up and the presence of the defects.



2. Tecnoss® Lamina® Hard is used to reconstruct the palatal and buccal walls in combination with Apatos® and autogenous bone chips.



3. After the treatment, the patients recovered her natural smile.

### HORIZONTAL/VERTICAL AUGMENTATION



## THE VESTIBULAR SOCKET THERAPY

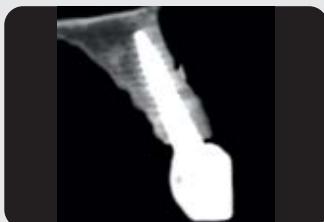
Sex: female - Age: 35 years old - Author: Dr. Abdelsalam Elaskary, private practice, Egypt



1. The CBCT scan shows a sagittal section of broken central incisor, with lost labial bone plate loss.



2. Lamina® Soft is stabilized using two membrane tacs apically according to the VST protocol.



3. The two years post-operative CBCT scan shows the regeneration of the labial plate of bone.

### ALVEOLAR REGENERATION



## TREATMENT OF A DEFECT IN THE ESTHETIC AREA

Sex: female - Age: 35 years old - Author: Prof. Tiziano Testori, University of Michigan, USA



1. The clinical pre-operative view shows a horizontal/vertical defect in the upper jaw.



2. Lamina® is used to protect the underlying collagenic biomaterial and regenerate the missing buccal bone.



3. Five-year follow-up. Soft tissues are healthy and the patient did not report any complication.

### HORIZONTAL/VERTICAL AUGMENTATION





## THE MODIFIED SHELL TECHNIQUE

Sex: female - Age: 28 years old - Author: Prof. Arndt Happe, private practice, Germany



1. The CBCT scan shows a Class V defect with missing palatal wall.



2. Tecnoss® Lamina® Hard is fixed buccally with osteosynthesis screws (1mm diameter) and the defect is filled with AB chips.



3. After four months, the clinical occlusal view shows a regenerated ridge.

## VERTICAL AUGMENTATION



## THE MULTI-LAYER TECHNIQUE

Sex: female - Age: 64 years old - Authors: Dr. Christian Maischberger and Prof. Hannes Wachtel, private practice, Germany



1. The clinical image shows an intraoperative occlusal view of the upper jaw.



2. Lamina® Soft, GTO®, and subepithelial connective tissue graft are used for simultaneous hard and soft tissue augmentation.



3. The five-year follow-up clinical view shows no sign of complications in the area of the transplanted cuspid.

## ALVEOLAR REGENERATION



## TREATMENT OF A KNIFE-EDGE RIDGE

Sex: male - Age: 61 years old - Author: Dr. Pavel Yaroshevich, private practice, Russia



1. The occlusal view of the posterior mandible reveals a knife edge ridge.



2. Gen-Os® is placed to fill the edentulous area for augmentation under a dry Lamina® Curved.

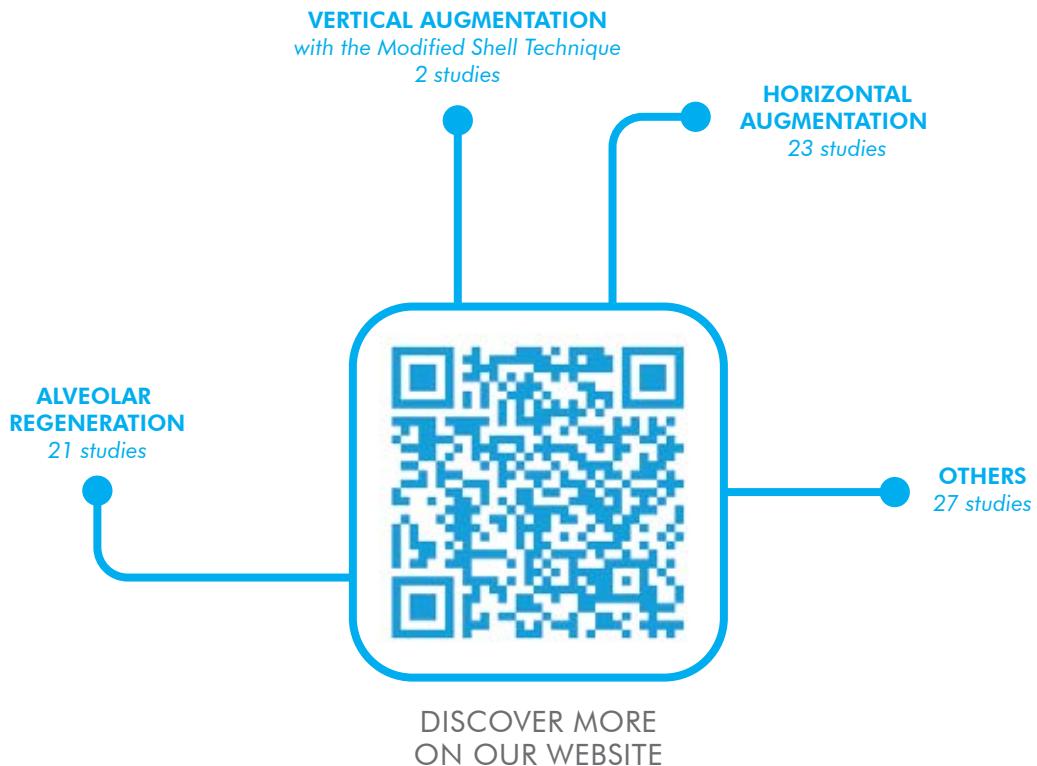


3. After six months following augmentation, an increase in the bone width on the buccal-lingual aspect and the bone normal quality is observed.

## HORIZONTAL/VERTICAL AUGMENTATION



# Lamina®'s scientific evidence



## SELECTED SCIENTIFIC BIBLIOGRAPHY

### ALVEOLAR REGENERATION

Festa VM, Addabbo F, Laino L, Femiano F, Rullo R. **Porcine-derived xenograft combined with a soft cortical membrane versus extraction alone for implant site development: a clinical study in humans.** Clin Implant Dent Relat Res. 2013 Oct;15(5):707-13

Elaskary A, Gaweesh YY, El Tantawi M, Maebed MA. **Vestibular Socket Therapy: a novel approach for implant placement in defective fresh extraction sockets with or without active socket infection (One-Arm Cohort Study).** Int J Oral Maxillofac Implants. 2021 Jan-Feb;36(1):146-153

Elaskary A, Meabed M, Abd-EIWahab Radi I. **Vestibular Socket Therapy with immediate implant placement for managing compromised fresh extraction sockets: a prospective single-arm clinical study.** Int J Oral Implantol (Berl). 2021 Aug 20;14(3):307-32

Schuh PL, Wachtel H, Beuer F, Goker F, Del Fabbro M, Francetti L, Testori T. **Multi-Layer Technique (MLT) with porcine collagenated cortical bone lamina for bone regeneration procedures and immediate post-extraction implantation in the esthetic area: a retrospective case series with a mean follow-up of 5 years.** Materials (Basel). 2021 Sep 9;14(18):5180

Ghalla NA, Elaskary A, Elsabagh H, Toukhy AE, Abdelrahman H, El-Kimary G.

**A novel atraumatic extraction technique using Vestibular Socket Therapy for immediate implant placement: a randomized controlled clinical trial.** Oral Maxillofac Surg. 2022 Jun 20

ElAskary A, Elfana A, Meabed M, Abd-EIWahab Radi I, Akram M, Fawzy El-Sayed K.

**Immediate implant placement utilizing Vestibular Socket Therapy versus early implant placement with contour augmentation for rehabilitation of compromised extraction sockets in the esthetic zone: a randomized controlled clinical trial.** Clin Implant Dent Relat Res. 2022 Oct;24(5):559-568

Körner G, Bäumer-König A, Barz-Popp F, Tripodakis A. **Esthetic implant solutions in the periodontally compromised anterior region. Expanding the indications for immediate implant placement with the Socket Rebuilding Technique (SRT).** Int J Esthet Dent. 2022 Nov 25;17(4):408-42

Elaskary A, Abdelrahman H, Elfahl B, Elsabagh H, El-Kimary G, Ghalla NA. **Immediate implant placement in intact fresh extraction sockets using Vestibular Socket Therapy versus partial extraction therapy in the esthetic zone: a randomized clinical trial.** Int J Oral Maxillofac Implants. 2023 May, Jun;38(3):468-478

Hamed MM, El-Tonsy MM, Elskary A, Abdelaziz GO, Saeed SS, Elfahl BN.  
**Effect of three different grafting materials on immediate implant placement using Vestibular Socket Therapy in Class II extraction sockets in the maxillary esthetic zone: a randomized controlled clinical trial.**  
BMC Oral Health. 2023 Sep 1;23(1):623

Elskary A, Ghallab N, Thabet A, Shemais N.  
**The bone shielding versus Dual-Zone concept in treating thin-walled fresh extraction sockets with immediate implant placement: soft and hard tissue changes. A randomized clinical trial.**  
Clin Implant Dent Relat Res. 2023 Sep 5

Köttgen C, Köttgen I, Plaster U.  
**Immediate implant placement in a dialysis patient. A case report.**  
Int J Esthet Dent. 2024 May 10;19(2):152-169

Elskary A, Thabet A, Hussin M, Radi IA.  
**Soft and hard tissue evaluation for Vestibular Socket Therapy of immediately placed implants in infected and non-infected sockets: a 1-year prospective cohort study.**  
BMC Oral Health. 2024 Oct 7;24(1):1190

#### EXPERIMENTAL STUDIES

Fischer KR, Götz W, Kauffmann F, Schmidlin PR, Friedmann A.  
**Ridge preservation of compromised extraction sockets applying a soft cortical membrane: a canine proof-of-principle evaluation.**  
Ann Anat. 2020 Sep;231:151524

#### HORIZONTAL AUGMENTATION

Wachtel H, Fickl S, Hinze M, Bolz W, Thalmair T.  
**The Bone Lamina Technique: a novel approach for lateral ridge augmentation. A case series.**  
Int J Periodontics Restorative Dent. 2013 Jul-Aug;33(4):491-7

Kivovics M, Foti V, Mayer Y, Mijiritsky E.  
**Fibrinogen-Induced Regeneration Sealing Technique (F.I.R.S.T.): a retrospective clinical study on 105 implants with a 3-7-year follow-up.**  
J Clin Med. 2024 Nov 17;13(22):6916

#### HORIZONTAL AND VERTICAL AUGMENTATION

Rossi R, Ghezzi C, Tomecek M.  
**Cortical Lamina: a new device for the treatment of moderate and severe tridimensional bone and soft tissue defects.**  
Int J Esthet Dent. 2020;15(4):454-473

#### IN VITRO STUDIES

Canullo L, Genova T, Naenni N, Nakajima Y, Masuda K, Mussano F.  
**Plasma of argon enhances the adhesion of murine osteoblasts on different graft materials.**  
Ann Anat. 2018 Jul;218:265-270

Di Carlo R, Zara S, Ventrella A, Siani G, Da Ros T, Iezzi G, Cataldi A, Fontana A.  
**Covalent decoration of cortical membranes with graphene oxide as a substrate for dental pulp stem cells.**  
Nanomaterials (Basel). 2019 Apr 12;9(4):604

Caballé-Serrano J, Munar-Frau A, Delgado L, Pérez R, Hernández-Alfaro F.  
**Physicochemical characterization of barrier membranes for bone regeneration.**  
J Mech Behav Biomed Mater. 2019 Sep;97:13-20

Radunovic M, Pavic A, Ivanovic V, Milivojevic M, Radovic I, Di Carlo R, Pilato S, Fontana A, Piattelli A, Petrovic S.  
**Biocompatibility and antibiofilm activity of graphene-oxide functionalized titanium discs and collagen membranes.**  
Dent Mater. 2022 Jul;38(7):1117-1127

#### ORBITAL FLOOR RESTORATION

Rinna C, Ungari C, Saltarel A, Cassoni A, Reale G.  
**Orbital floor restoration.**  
J Craniofac Surg. 2005 Nov;16(6):968-72

Ozel B, Findikcioglu K, Sezgin B, Guney K, Barut I, Ozmen S.  
**A new option for the reconstruction of orbital floor defects with heterologous cortical bone.**  
J Craniomaxillofac Surg. 2015 Oct;43(8):1583-8

Senese O, Bourtremans E, Gossiaux C, Loeb I, Dequanter D.  
**Retrospective analysis of 79 patients with orbital floor fracture: outcomes and patient-reported satisfaction.**  
Arch Craniofac Surg. 2018 Jun;19(2):108-113

Aboh IV, Chisci G, Gennaro P, Giovannetti F, Bartoli D, Priore P, Anelli A, Iannetti G.  
**Complications of orbital floor repair with silastic sheet: the skin fistula.**  
J Craniofac Surg. 2013 Jul;24(4):1288-91

#### REVIEWS

Rondone EM, Leitão-Almeida B, Pereira MS, Fernandes GVO, Borges T.  
**The use of tissue grafts associated with immediate implant placement to achieve better peri-implant stability and efficacy: a systematic review and meta-analysis.**  
J Clin Med. 2024 Jan 3; 1;13(3):821

#### VERTICAL AUGMENTATION

Villa G, Rizzacasa A, Bessa L, Spina P, Barootchi S, Tavelli L, Wang HL.  
**Shell Technique with a xenogeneic cortical bone lamina and particulate bone graft for horizontal ridge augmentation: a case series.**  
Int J Periodontics Restorative Dent. 2023 July-Aug;43(4):435-441

Happe A, Blender SM, Luthardt RG, Rudolph H, Kuhn K.  
**Digital evaluation of vertical ridge augmentation with the Modified Shell Technique using a xenogeneic bone lamina: a case series.**  
Journal of Clinical Medicine. 2023; 12(22):7013

# **Evolution**

***The natural Evolution  
of collagen membranes***

# Why choose **Evolution**?

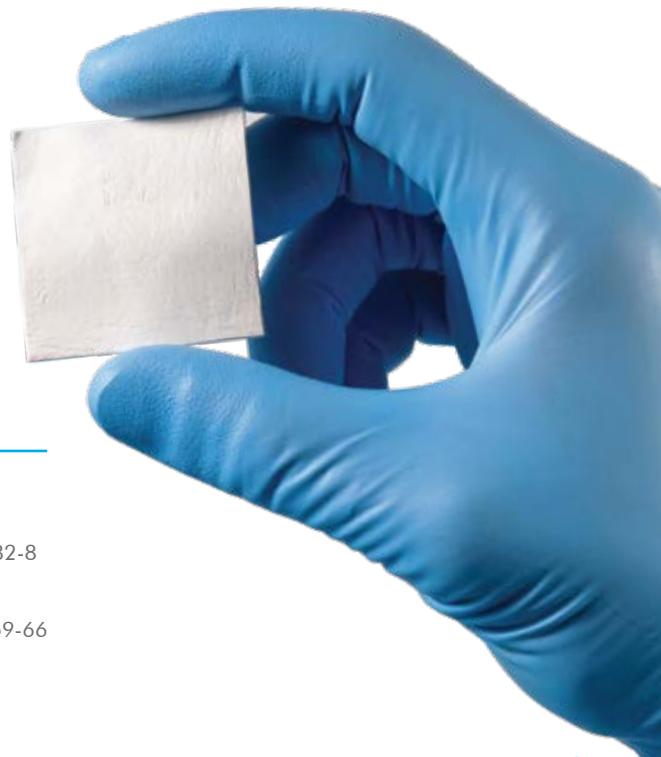
---

Graft protection against bacteria and food contamination is necessary to allow hard and soft tissue healing. Thanks to their high consistency, resistance to degradation<sup>(1)</sup> and stretchability, **Evolution** collagen membranes are valid solutions to maintain the bone graft stable and isolated thus reducing the risk of complications<sup>(2-7)</sup>.

The structure made of dense collagen fibres<sup>(8)</sup> and the appropriate gradual degradation rate prevents the infection from bacteria and soft tissue ingrowth.

In addition, when exposed intentionally or unintentionally, **Evolution** collagen membranes allow second-intention soft tissue healing<sup>(3)</sup> and adequate new bone formation<sup>(2-7, 9,10)</sup>.

More than 25 years of clinical and scientific evidence prove that **Evolution** collagen membranes can be used in post-extraction sockets to protect the graft<sup>(2,3)</sup>, in lateral access sinus lift to protect both the antrostomy and the sinus membrane<sup>(4)</sup>, to stabilise the graft around peri-implant lesions<sup>(5)</sup>, periodontal defects<sup>(6)</sup>, and vertical defects<sup>(7)</sup>.



---

1. Vallecillo-Rivas M et al. Polymers (Basel), 2021 Sep 5;13(17):3007
2. Barone A et al. Clin Oral Implants Res, 2015 Jul;26(7):806-13
3. Barone A et al. J Periodontol, 2014 Jan;85(1):14-23
4. Cassetta M et al. Int J Periodontics Restorative Dent. 2012 Dec;32(6):e182-8
5. Barone A et al. Clin Implant Dent Relat Res, 2016 Dec;18(6):1103-1112
6. Esposito et al. Eur J Oral Implantol, 2015 Autumn;8(3):233-44
7. Felice P et al. Int J Periodontics Restorative Dent, 2013 Mar-Apr;33(2):159-66
8. Osorio R et al. Polymers (Basel), 2022 Aug 11;14(16):3274
9. Iida T et al. Clin Oral Implants Res, 2017 Dec;28(12):1567-1576
10. Hirota A et al. Int J Implant Dent, 2019 Aug 19;5(1):31

# Clinical cases & applications

## TREATMENT OF A FRACTURED TOOTH

Sex: male - Age: 53 years old - Author: Prof. Antonio Barone, University of Pisa, Italy



1. The periapical x-ray shows a hopeless tooth.



2. The fresh extraction socket is grafted with mp3® and covered with an *Evolution* collagen membrane.



3. The x-ray at three-year follow-up shows an implant surrounded by appropriate amount of newly formed bone.

### ALVEOLAR REGENERATION



## THE "ALL-ON-4" PROTOCOL FOR TRANS-SINUS IMPLANTATION

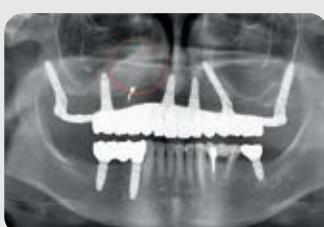
Sex: female - Age: 62 years old - Author: Prof. Aleksei Reshetnikov, private practice, Russia



1. Panoramic radiograph of the bone tissue. The red circle shows the area with the most pronounced atrophy of the alveolar ridge of the upper jaw.



2. The oroantral anastomosis is isolated with an *Evolution* collagen membrane. The antrostomy is filled with mp3®.



3. Panoramic radiograph six months after implantation shows a permanent prosthesis installed on the upper jaw.

### SINUS MEMBRANE PROTECTION



## IMPLANT PLACEMENT IN ESTHETIC REGION AND RECONSTRUCTION OF THE BONE DEFECT

Sex: female - Age: 33 years old - Author: Dr. Matej Kuliš, private practice, Slovenia



1. After extraction and removal of the granulation tissue, a buccal bone defect is reported.

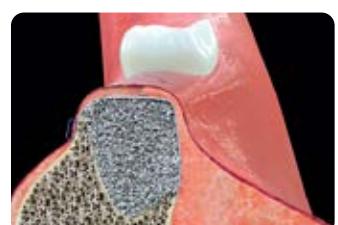


2. The *Evolution* collagen membrane is adapted to the defect anatomy to cover and stabilise the underlying graft.



3. The x-ray taken after second stage surgery reveals an appropriate amount of newly formed bone.

### ALVEOLAR REGENERATION





## REPLACEMENT OF A FRACTURED TOOTH

Sex: female - Age: 35 years old - Author: Dr. Guerino Paolantoni, private practice, Italy



1. The x-ray shows the upper left incisor with a horizontal fracture of the root.



2. An Evolution collagen membrane is stabilised with titanium pins to cover the graft.



3. The x-ray shows an implant surrounded by healthy new bone.

### ALVEOLAR REGENERATION



## THE "ALL-ON-4" PROTOCOL

Sex: male - Age: 37 years old - Author: Prof. Aleksei Reshetnikov, private practice, Russia



1. After teeth extraction, the lower jaw is ready for implant insertion.



2. mp3® is packed around the installed implants and Evolution collagen membranes cover alveolar ridge defects.

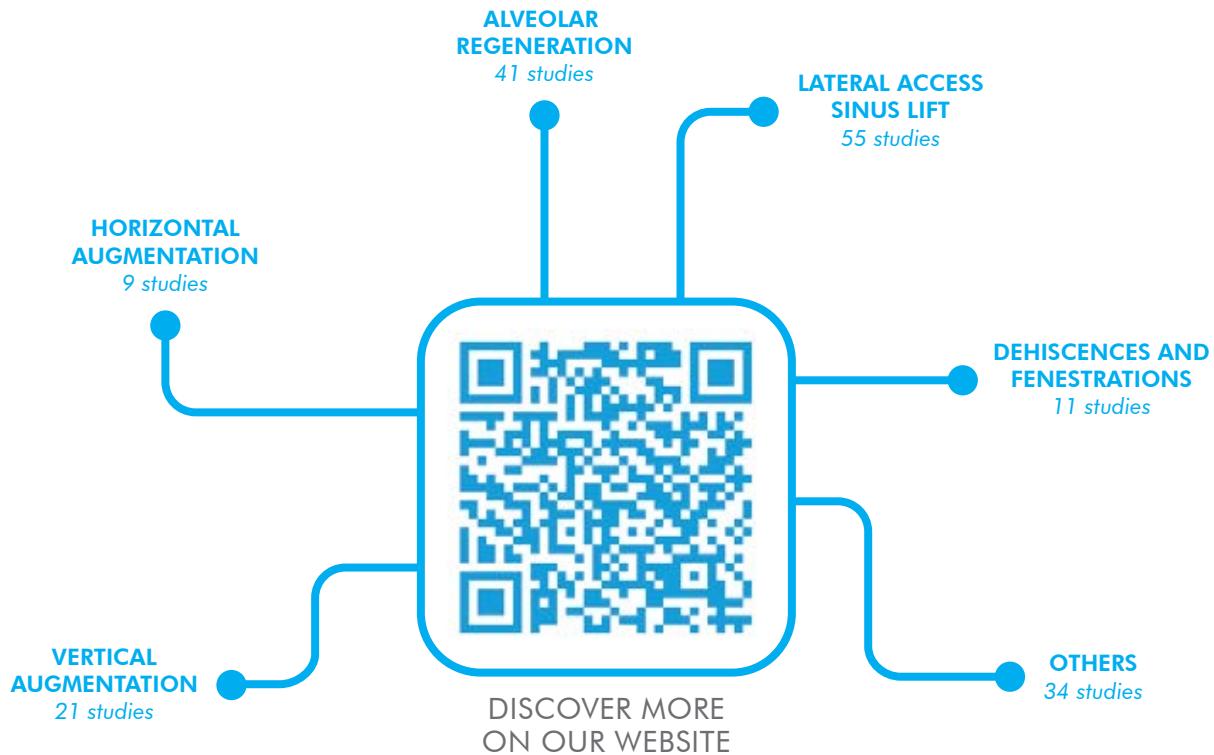


3. Six months after the surgery, the soft tissue is healthy.

### ALVEOLAR REGENERATION



# Evolution's scientific evidence



## SELECTED SCIENTIFIC BIBLIOGRAPHY

### ALVEOLAR REGENERATION

Barone A, Aldini NN, Fini M, Giardino R, Calvo Guirado JL, Covani U. **Xenograft versus extraction alone for ridge preservation after tooth removal: a clinical and histomorphometric study.** J Periodontol. 2008 Aug;79(8):1370-7

Cardaropoli D, Cardaropoli G. **Preservation of the postextraction alveolar ridge: a clinical and histologic study.** Int J Periodontics Restorative Dent. 2008 Oct;28(5):469-77

Barone A, Orlando B, Cingano L, Marconcini S, Derchi G, Covani U. **A randomized clinical trial to evaluate and compare implants placed in augmented versus non-augmented extraction sockets: 3-year results.** J Periodontol. 2012 Jul;83(7):836-46

Barone A, Ricci M, Tonelli P, Santini S, Covani U. **Tissue changes of extraction sockets in humans: a comparison of spontaneous healing vs. ridge preservation with secondary soft tissue healing.** Clin Oral Implants Res. 2013 Nov;24(11):1231-7

Barone A, Borgia V, Covani U, Ricci M, Piattelli A, Iezzi G. **Flap versus flapless procedure for ridge preservation in alveolar extraction sockets: a histological evaluation in a randomized clinical trial.** Clin Oral Implants Res. 2015 Jul;26(7):806-13

Barone A, Ricci M, Romanos GE, Tonelli P, Alfonsi F, Covani U. **Buccal bone deficiency in fresh extraction sockets: a prospective single cohort study.** Clin Oral Implants Res. 2015 Jul;26(7):823-30

Barone A, Toti P, Quaranta A, Alfonsi F, Cucchi A, Calvo-Guirado JL, Negri B, Di Felice R, Covani U. **Volumetric analysis of remodelling pattern after ridge preservation comparing use of two types of xenografts. A multicentre randomized clinical trial.** Clin Oral Implants Res. 2016 Nov;27(11):e105-e115

Barone A, Toti P, Quaranta A, Alfonsi F, Cucchi A, Negri B, Di Felice R, Marchionni S, Calvo-Guirado JL, Covani U, Nannmark U. **Clinical and histological changes after ridge preservation with two xenografts: preliminary results from a multicentre randomized controlled clinical trial.** J Clin Periodontol. 2017 Feb;44(2):204-214

Giuliani A, Iezzi G, Mazzoni S, Piattelli A, Perrotti V, Barone A. **Regenerative properties of collagenated porcine bone grafts in human maxilla: demonstrative study of the kinetics by synchrotron radiation microtomography and light microscopy.** Clin Oral Investig. 2018 Jan;22(1):505-513

Checchi V, Felice P, Zucchelli G, Barausse C, Piattelli M, Pistilli R, Grandi G, Esposito M. **Wide diameter immediate post-extractive implants vs delayed placement of normal-diameter implants in preserved sockets in the molar region: 1-year post-loading outcome of a randomised controlled trial.** Eur J Oral Implantol. 2017;10(3):263-278

Kilinc A, Ataol M.

**How effective is collagen resorbable membrane placement after partially impacted mandibular third molar surgery on postoperative morbidity? A prospective randomized comparative study.**

BMC Oral Health. 2017 Oct 5;17(1):126

Barone A, Toti P, Piattelli A, Iezzi G, Derchi G, Covani U. **Extraction socket healing in humans after ridge preservation techniques: comparison between flapless and flapped procedures in a randomized clinical trial.** J Periodontol. 2014 Jan;85(1):14-23

Marconcini S, Giammarinaro E, Derchi G, Alfonsi F, Covani U, Barone A.

**Clinical outcomes of implants placed in ridge-preserved versus nonpreserved sites: a 4-year randomized clinical trial.**

Clin Implant Dent Relat Res. 2018 Dec;20(6):906-914 294

Kilinc A, Ataol M.

**Effects of collagen membrane on bone level and periodontal status of adjacent tooth after third molar surgery: a randomized controlled trial.**

Head Face Med. 2023 Mar 25;19(1):13

#### DEHISCENCE AND FENESTRATIONS

Covani U, Cornelini R, Barone A.

**Buccal bone augmentation around immediate implants with and without flap elevation: a modified approach.**

Int J Oral Maxillofac Implants. 2008 Sep-Oct;23(5):841-6

Covani U, Canullo L, Toti P, Alfonsi F, Barone A.

**Tissue stability of implants placed in fresh extraction sockets: a 5-year prospective single-cohort study.**

J Periodontol. 2014 Sep;85(9):e323-32

#### HORIZONTAL AND VERTICAL AUGMENTATION

Wachtel H, Fickl S, Hinze M, Bolz W, Thalmair T.

**The Bone Lamina Technique: a novel approach for lateral ridge augmentation: a case series.**

Int J Periodontics Restorative Dent. 2013 Jul-Aug;33(4):491-7

Felice P, Piana L, Checchi L, Corvino V, Nannmark U, Piattelli M. **Vertical ridge augmentation of an atrophic posterior mandible with an inlay technique and cancellous equine bone block: a case report.**

Int J Periodontics Restorative Dent. 2013 Mar-Apr;33(2):159-66

Felice P, Barausse C, Barone A, Zucchelli G, Piattelli M, Pistilli R, Ippolito DR, Simion M.

**Interpositional augmentation technique in the treatment of posterior mandibular atrophies: a retrospective study comparing 129 autogenous and heterologous bone blocks with 2 to 7 years follow-up.**

Int J Periodontics Restorative Dent. 2017 Jul/Aug;37(4):469-480

Happe A, Blender SM, Luthardt RG, Rudolph H, Kuhn K. **Digital evaluation of vertical ridge augmentation with the Modified Shell Technique using a xenogeneic bone Lamina: a case series.**

J Clin Med. 2023 Nov 9;12(22):7013

#### LATERAL ACCESS SINUS LIFT

Barone A, Crespi R, Aldini NN, Fini M, Giardino R, Covani U. **Maxillary sinus augmentation: histologic and histomorphometric analysis.**

Int J Oral Maxillofac Implants. 2005 Jul-Aug;20(4):519-25

Barone A, Santini S, Sbordone L, Crespi R, Covani U.

**A clinical study of the outcomes and complications associated with maxillary sinus augmentation.**

Int J Oral Maxillofac Implants. 2006 Jan-Feb;21(1):81-5

Scarano A, Piattelli A, Assenza B, Quaranta A, Perrotti V, Piattelli M, Iezzi G.

**Porcine bone used in sinus augmentation procedures: a 5-year retrospective clinical evaluation.**

J Oral Maxillofac Surg. 2010 Aug;68(8):1869-73

Scarano A, Piattelli A, Perrotti V, Manzon L, Iezzi G.

**Maxillary sinus augmentation in humans using cortical porcine bone: a histological and histomorphometrical evaluation after 4 and 6 months.**

Clin Implant Dent Relat Res. 2011 Mar;13(1):13-8

Ramírez-Fernández MP, Calvo-Guirado JL, Maté-Sánchez Del Val JE, Delgado-Ruiz RA, Negri B, Barona-Dorado C. **Ultrastructural study by backscattered electron imaging and elemental microanalysis of bone-to-biomaterial interface and mineral degradation of porcine xenografts used in maxillary sinus floor elevation.**

Clin Oral Implants Res. 2013 May;24(5):523-30

Tanaka K, Iezzi G, Piattelli A, Ferri M, Mesa NF, Apaza Alccayhuaman KA, Botticelli D.

**Sinus floor elevation and antrostomy healing: a histomorphometric clinical study in humans.**

Implant Dent. 2019 Dec;28(6):537-542

Pistilli R, Canullo L, Pesce P, Pistilli V, Caponio VCA, Sbricoli L **Guided implant surgery and sinus lift in severely resorbed maxillae: a retrospective clinical study with up to 10 years of follow-up.**

J Dent. 2022 Jun;121:104137

Morimoto A, Kobayashi N, Ferri M, Iezzi G, Piattelli A, Fortich Mesa N, Botticelli D.

**Influence on implant bone healing of a collagen membrane placed subjacent the sinus mucosa - A randomized clinical trial on sinus floor elevation.**

Dent J (Basel). 2022 Jun 8;10(6):105

Imai H, Iezzi G, Piattelli A, Ferri M, Apaza Alccayhuaman KA, Botticelli D.

**Influence of the dimensions of the antrostomy on osseointegration of mini-implants placed in the grafted region after sinus floor elevation: a randomized clinical trial.**

Int J Oral Maxillofac Implants. 2020 May/Jun;35(3):591-598

Hirota A, Iezzi G, Piattelli A, Ferri M, Tanaka K, Apaza Alccayhuaman KA, Botticelli D.

**Influence of the position of the antrostomy in sinus floor elevation on the healing of mini-implants: a randomized clinical trial.**

Oral Maxillofac Surg. 2020 Sep;24(3):299-308

Correia F, Gouveia S, Felino AC, Faria-Almeida R, Pozza DH. **Maxillary sinus augmentation with xenogenic collagen-retained heterologous cortico-cancellous bone: a 3-year follow-up randomized controlled trial.**

Dentistry Journal. 2024; 12(2):33

#### PERIODONTAL REGENERATION

Cardaropoli D, Cardaropoli G.

**Healing of gingival recessions using a collagen membrane with a hemineralized xenograft: a randomized controlled clinical trial.**

Int J Periodontics Restorative Dent. 2009 Feb;29(1):59-67

Esposito M, Grusovin MG, Lambert F, Matos S, Pietruska M, Rossi R, Salhi L, Buti J.

**The effectiveness of a resorbable bone substitute with a resorbable membrane in the treatment of periodontal infrabony defect - A multicenter randomised controlled trial.**

Eur J Oral Implantol. 2015 Autumn;8(3):233-44

# **Derma**

***The versatile  
collagen membrane***

# Why choose **Derma**?

---

Connective Tissue Graft (CTG) is considered the gold standard for soft tissue regeneration; however, autologous soft tissue is available in limited quantities via palatal harvesting and causes post-operative discomfort to patients.

**Derma** is a valid alternative to CTG for soft tissue augmentation around teeth and implants and for the treatment of gingival recessions<sup>(1-4)</sup>.

Natural dense collagen fibres make **Derma** a resistant membrane also for sealing the socket and stabilising the graft in alveolar regeneration procedures<sup>(4-6)</sup> where it can be left exposed without any risk of infection<sup>(4-6)</sup>.



---

1. Matoh U et al. Int J Periodontics Restorative Dent, 2019;39(3):439-446
2. Fischer K et al. Clin Implant Dent Relat Res, 2019;1-8
3. Fischer K et al. Quintessence Int, 2014 Nov-Dec;45(10):853-60
4. Minniti A et al. J Clin Med, 2024 May 17;13(10):2954
5. Tallarico M et al. Eur J Oral Implantol, 2016;9(3):263-27
6. Tallarico M et al. Eur J Oral Implantol, 2017;10(2):169-178

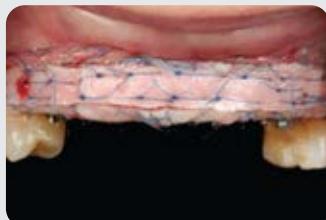
# Clinical cases & applications

## SOFT TISSUE REGENERATION IN THE ESTHETIC AREA

Sex: female - Age: 53 years old - Author: Dr. Iñaki Gamborena, private practice, Spain



1. The patient lost her natural smile.



2. The strip gingival autograft technique is selected in order to blend the gingival color with the rest of the soft tissues in conjunction with Derma.



3. After the treatment, the patient recovered a natural smile.

### SOFT TISSUE AUGMENTATION



## SEALING OF A LOWER MOLAR POST-EXTRACTION SOCKET

Sex: female - Age: 42 years old - Author: Dr. Domenico Patarino, private practice, Italy



1. The clinical image shows a post-extractive molar socket.



2. Derma is used to protect the underlying Gen-Os® bone graft.



3. Derma did not get infected and favoured soft tissue healing.

### ALVEOLAR REGENERATION



## THE MATRIX TISSUE GRAFT (MTG) TECHNIQUE

Sex: female - Age: 37 years old - Author: Dr. Alessandro Minniti, private practice, Italy



1. The initial situation shows a buccal and lingual hard and soft tissue atrophy.



2. Derma is inserted buccally to augment the soft tissue.



3. A sufficient tissue thickness is gained for a natural soft tissue emergence profile.

### SOFT TISSUE AUGMENTATION





## TREATMENT OF SEVERE GINGIVAL RECESION

Sex: female - Age: 55 - Author: Prof. Rok Gašperšič, University of Lubljana, Slovenia



1. The patient shows a severe gingival recession.



2. Derma is sutured with mattress sutures.



3. Two years after surgery, the patient shows a normal soft tissue profile.

### SOFT TISSUE AUGMENTATION

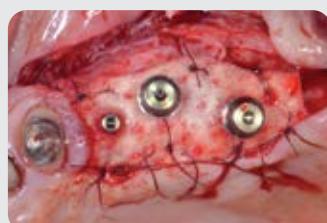


## SINUS LIFT COMBINED WITH ALVEOLAR RECONSTRUCTION

Sex: female - Age: 67 - Author: Dr Ferdinando D'Avenia, private practice, Italy



1. Clinical view before soft tissue management.

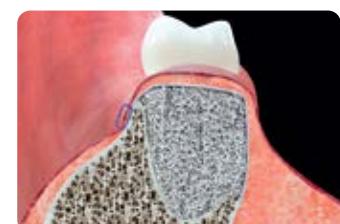


2. Derma graft is used for connective tissue protection.

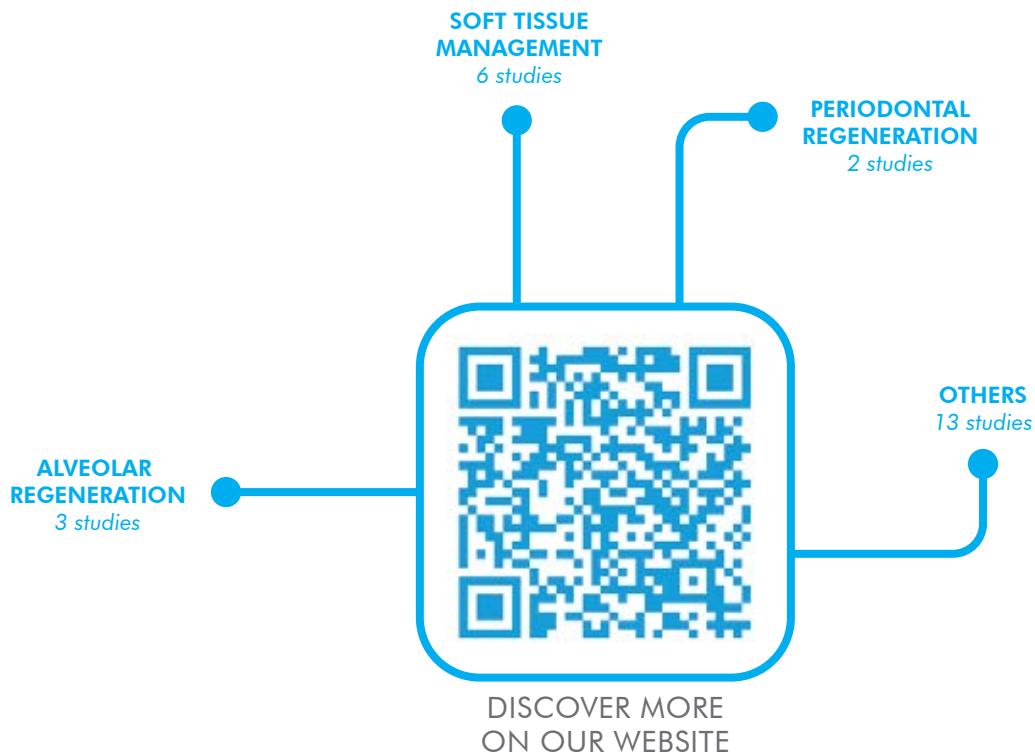


3. Eight weeks healing, at the time of provisional restoration application.

### SOFT TISSUE AUGMENTATION



# Derma's scientific evidence



## SELECTED SCIENTIFIC BIBLIOGRAPHY

### ALVEOLAR REGENERATION

Tallarico M, Xhanari E, Pisano M, De Riu G, Tullio A, Meloni SM. Single post-extractive ultra-wide 7 mm-diameter implants versus implants placed in molar healed sites after socket preservation for molar replacement: 6-month post-loading results from a randomised controlled trial. *Eur J Oral Implantol.* 2016;9(3):263-275

Ramanauskaite A, Borges T, Almeida BL, Correia A. Dental implant outcomes in grafted sockets: a systematic review and meta-analysis. *J Oral Maxillofac Res.* 2019 Sep 5;10(3):e8

Tallarico M, Xhanari E, Pisano M, Gatti F, Meloni SM. Molar replacement with 7 mm-wide diameter implants: to place the implant immediately or to wait 4 months after socket preservation? 1 year after loading results from a randomised controlled trial. *Eur J Oral Implantol.* 2017;10(2):169-178

### EXPERIMENTAL STUDIES

Fickl S, Nannmark U, Schlagenhauf U, Hürzeler MB, Kehschull M. Porcine dermal matrix in the treatment of dehiscence-type defects-an experimental split-mouth animal trial. *Clin Oral Implants Res.* 2015 Jul;26(7):799-805

Fischer KR, Götz W, Kauffmann F, Schmidlin PR, Friedmann A. Ridge preservation of compromised extraction sockets applying a soft cortical membrane: a canine proof-of-principle evaluation. *Ann Anat.* 2020 Sep;231:151524

Aragoneses J, Suárez A, Rodríguez C, Aragoneses JM. Clinical and histological differences between guided tissue regeneration with acellular dermal matrix of porcine origin and autologous connective tissue: an animal study. *Materials (Basel).* 2021 Jan 7;14(2):27

### IN VITRO STUDIES

De Marco P, Zara S, De Colli M, Radunovic M, Lazović V, Ettorre V, Di Crescenzo A, Piattelli A, Cataldi A, Fontana A. Graphene oxide improves the biocompatibility of collagen membranes in an in vitro model of human primary gingival fibroblasts. *Biomed Mater.* 2017 Sep 13;12(5):055005

Radunovic M, De Colli M, De Marco P, Di Nisio C, Fontana A, Piattelli A, Cataldi A, Zara S. Graphene oxide enrichment of collagen membranes improves DPSCs differentiation and controls inflammation occurrence. *J Biomed Mater Res A.* 2017 Aug;105(8):2312-2320

Toledano M, Asady S, Toledano-Osorio M, García-Godoy F, Serrera-Figallo MA, Benítez-García JA, Osorio R. Differential biodegradation kinetics of collagen membranes for bone regeneration. *Polymers (Basel).* 2020 Jun 4;12(6):1290

Azab E, Youssef AR. Biocompatibility evaluation of human and porcine acellular dermal matrix on human primary gingival fibroblasts: in vitro comparative study. *Eur J Dent.* 2021 Jul;15(3):563-567

Osorio R, Asady S, Toledano-Osorio M, Toledano M, Bueno JM, Martínez-Ojeda RM, Osorio E.  
**Biomimetic remineralization of an extracellular matrix collagen membrane for bone regeneration.**  
Polymers (Basel). 2022 Aug 11;14(16):3274

Lazarevic M, Petrovic S, Pierfelice TV, Ignjatovic N, Piattelli A, Vlajic Tovilovic T, Radunovic M.  
**Antimicrobial and osteogenic effects of collagen membrane decorated with chitosan-nano-hydroxyapatite.**  
Biomolecules. 2023 Mar 23;13(4):579

#### PERIODONTAL REGENERATION

Fickl S, Jockel-Schneider Y, Lincke T, Bechtold M, Fischer KR, Schlagenauf U.  
**Porcine dermal matrix for covering of recession type defects: a case series.**  
Quintessence Int. 2013 Mar;44(3):243-6

Matoh U, Petelin M, Gašperšič R.  
**Split-mouth comparison of coronally advanced flap with connective tissue graft or collagen matrix for treatment of isolated gingival recessions.**  
Int J Periodontics Restorative Dent. 2019 May/Jun;39(3):439-446

#### REVIEWS

Romasco T, Tumedei M, Inchingolo F, Pignatelli P, Montesani L, Iezzi G, Petrini M, Piattelli A, Di Pietro N.  
**A narrative review on the effectiveness of bone regeneration procedures with OsteoBiol® collagenated porcine grafts: the translational research experience over 20 years.**  
J Funct Biomater. 2022 Aug 18;13(3):121

#### SOFT TISSUE MANAGEMENT

Fischer KR, Testori T, Wachtel H, Mühlmann S, Happe A, Del Fabbro M.  
**Soft tissue augmentation applying a collagenated porcine dermal matrix during second stage surgery: a prospective multicenter case series.**  
Clin Implant Dent Relat Res. 2019 Oct;21(5):923-930

Fischer KR, Fickl S, Mardas N, Bozec L, Donos N.  
**Stage-two surgery using collagen soft tissue grafts: clinical cases and ultrastructural analysis.**  
Quintessence Int. 2014 Nov-Dec;45(10):853-60

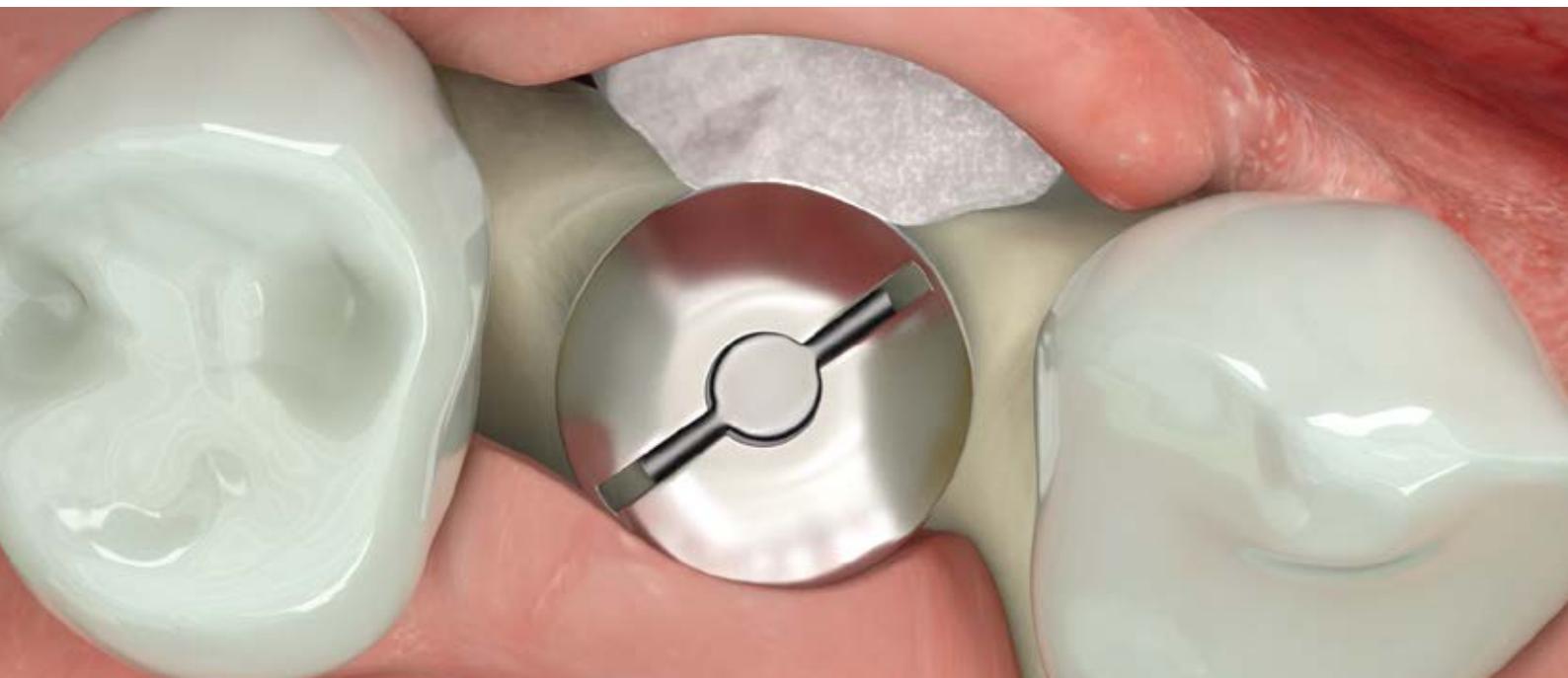
Verardi S, Orsini M, Lombardi T, Ausenda F, Testori T, Pulici A, Oreglia F, Valente NA, Stacchi C.  
**Comparison between two different techniques for peri-implant soft tissue augmentation: porcine dermal matrix graft versus tenting screw.**  
J Periodontol. 2020 Aug;91(8):1011-1017

Minniti A, Caroprese M, Zarantonello M, De Santis D, Caliendo G, Gelpi F.  
**High-density dermal matrix for soft tissue augmentation using a Matrix Tissue Graft Technique a comprehensive multicenter analysis of 20 implants: a 1-year follow-up retrospective study.**  
J Clin Med. 2024 May 17;13(10):2954

# OsteoBiol® Derma

by Tecnoss

PORCINE DERMAL MATRIX  
Natural xenogenic origin



#### Clinical evidence:

- **Horizontal soft tissue augmentation around implants**
- **As an alternative to connective tissue graft to improve the quality of keratinized tissues**
- **As prolonged protection of the underlying graft in socket regeneration procedures and in open healing situations**

# Our history



# Quality system

## ALWAYS FROM THE PATIENT'S SIDE

Tecnoss®, the Manufacturer of OsteoBiol® biomaterials, meets strict regulations that guarantee the safety of the OsteoBiol® product line.

First and foremost, the OsteoBiol® product line received the CE Certification, which guarantees the respect of high standards of safety: the conformity to such regulations is annually checked by Notified Bodies. OsteoBiol® product line meets the high-quality standards of biocompatibility as defined by ISO 10993.

OsteoBiol® collagenic xenografts are also sterilized according to international standard procedures such as EN ISO 11137-1, EN ISO 11137-2, and EN 556-1.

Product safety is also guaranteed by Tecnoss® risk management system (ISO 14971) and the quality management system is certified under EN ISO 13485.

Finally, more than 1 million surgeries, 350 scientific papers, and 25 years of clinical experience demonstrated OsteoBiol® product line is safe and trustworthy.

## A CHOICE THAT MATTERS

OsteoBiol® heterologous biomaterials are porcine or equine xenografts. For this reason, they are free from BSE (Bovine Spongiform Encephalopathy) and TSE (Transmissible Spongiform Encephalopathy) diseases. They are classified as "tissues with no detectable infectivity" (EN ISO 22442-1, and EN ISO 22442-3).

Raw materials are sourced from certified suppliers, where animals are checked by the local Veterinary Service. Furthermore, the OsteoBiol® product line is free from *Taenia solium* eggs/larvae since Tecnoss® processes only bone and mesenchymal tissue, whereas only striated muscles from the heart, tongue, and intercostal muscles are reservoirs of the aforementioned parasites.



# Our official distribution





**WE ARE WORLDWIDE**  
*Find your dealer*

# Our international courses

Each year we plan regeneration courses with top international speakers in exclusive locations.

Join the OsteoBiol® community to enhance your knowledge on our biomaterials and to experience their application in hands-on sessions.



[JOIN OUR COURSES](#)





# **Bone, Biomaterials & Beyond**

## Academy

Bone, Biomaterials & Beyond Academy is an international non-profit scientific association.

The Academy goal is to favor and disseminate the study of collagenic "Dual-Phase" bone substitutes of porcine or equine origin, and to investigate their clinical efficacy and predictability in oral surgery.

These goals are pursued by sustaining scientific research via professional courses, symposia and congresses, by organising contests, prizes and promoting scientific publications and any other initiative contributing to scientific progress.

CONGRESSES

CONTESTS

WEBINARS

Scan for more information about  
BBB Academy activities



# Bone Substitutes

## GTO®



Codes	Size	Origin	Type	Granulometry	Mean observed re-entry time
MU0005S	0,5 cc	Porcine	Bone Mix+TSV Gel	0,6-1 mm	About 5 months
MU0020S	2 cc	Porcine	Bone Mix+TSV Gel Wide tip syringe	0,6-1 mm	About 5 months
MU0005E	0,5 cc	Equine	Bone Mix+TSV Gel	0,6-1 mm	About 5 months
MU0020E	2 cc	Equine	Bone Mix+TSV Gel Wide tip syringe	0,6-1 mm	About 5 months

## mp3®



Codes	Size	Origin	Type	Granulometry	Mean observed re-entry time
A3095FS	0,5 cc	Porcine	Bone Mix	0,6-1 mm	About 5 months
A3005FS	1 cc	Porcine	Bone Mix	0,6-1 mm	About 5 months
A3010FS	2 cc	Porcine	Bone Mix Wide-tip syringe	0,6-1 mm	About 5 months
A3210FS	2 cc	Porcine	Bone Mix Wide-tip syringe	1-2 mm	About 5 months
A3095FE	0,5 cc	Equine	Bone Mix	0,6-1 mm	About 5 months
A3005FE	1 cc	Equine	Bone Mix	0,6-1 mm	About 5 months
A3010FE	2 cc	Equine	Bone Mix Wide-tip syringe	0,6-1 mm	About 5 months
A3210FE	2 cc	Equine	Bone Mix Wide-tip syringe	1-2 mm	About 5 months

## Gel 40



Codes	Size	Origin	Type	Granulometry	Mean observed re-entry time
05GEL40S	0,5 cc	Porcine	Bone Gel	≤ 0,300 mm	About 4 months

## Putty



Codes	Size	Origin	Type	Granulometry	Mean observed re-entry time
HPT52S	0,25 cc	Porcine	Bone Paste	≤ 0,300 mm	About 4 months
HPT09S	0,5 cc	Porcine	Bone Paste	≤ 0,300 mm	About 4 months
HPT61S	1 cc	Porcine	Bone Paste Wide-tip syringe	≤ 0,300 mm	About 4 months
HPT52E	0,25 cc	Equine	Bone Paste	≤ 0,300 mm	About 4 months
HPT09E	0,5 cc	Equine	Bone Paste	≤ 0,300 mm	About 4 months
HPT61E	1 cc	Equine	Bone Paste Wide-tip syringe	≤ 0,300 mm	About 4 months

## Gen-Os®



Codes	Size	Origin	Type	Granulometry	Mean observed re-entry time
M1052FS	0,25 g	Porcine	Dried Granules	0,25-1 mm	About 5 months
M1005FS	0,5 g	Porcine	Dried Granules	0,25-1 mm	About 5 months
M1010FS	1 g	Porcine	Dried Granules	0,25-1 mm	About 5 months
M1020FS	2 g	Porcine	Dried Granules	0,25-1 mm	About 5 months
M0210FS	1 g	Porcine	Dried Granules	1-2 mm	About 5 months
M0220FS	2 g	Porcine	Dried Granules	1-2 mm	About 5 months
M1052FE	0,25 g	Equine	Dried Granules	0,25-1 mm	About 5 months
M1005FE	0,5 g	Equine	Dried Granules	0,25-1 mm	About 5 months
M1010FE	1 g	Equine	Dried Granules	0,25-1 mm	About 5 months
M1020FE	2 g	Equine	Dried Granules	0,25-1 mm	About 5 months

## Apatos®



Codes	Size	Origin	Type	Granulometry	Mean observed re-entry time
A1005FS	0,5 g	Porcine	Dried Granules Mix	0,6-1 mm	About 5 months
A1010FS	1 g	Porcine	Dried Granules Mix	0,6-1 mm	About 5 months
A1020FS	2 g	Porcine	Dried Granules Mix	0,6-1 mm	About 5 months
A0210FS	1 g	Porcine	Dried Granules Mix	1-2 mm	About 5 months
AC1005FS	0,5 g	Porcine	Dried Granules Cortical	0,6-1 mm	About 5 months
AC1010FS	1 g	Porcine	Dried Granules Cortical	0,6-1 mm	About 5 months
A1005FE	0,5 g	Equine	Dried Granules Mix	0,6-1 mm	About 5 months
A1010FE	1 g	Equine	Dried Granules Mix	0,6-1 mm	About 5 months
A1020FE	2 g	Equine	Dried Granules Mix	0,6-1 mm	About 5 months
A0210FE	1 g	Equine	Dried Granules Mix	1-2 mm	About 5 months

# Bone Substitutes

Codes	Size	Origin	Type	Mean observed re-entry time
TSV005S	0,5 cc	Porcine	Bone Gel	About 5 months

**TSV Gel**



Codes	Size	Origin	Type	Mean observed re-entry time
BN0E	10x10x10 mm	Equine	Dried Block	About 8 months
BN1E	10x10x20 mm	Equine	Dried Block	About 8 months
BN2E	10x20x20 mm	Equine	Dried Block	About 8 months
BN8E	35x10x5 mm	Equine	Dried Block	About 8 months

**Sp-Block**



Codes	Size	Origin	Type	Thickness	Mean observed re-entry time
LS10HS	35x35 mm (Curved)	Porcine	Dried/Soft	1 mm	About 6 months
LS23FS	25x35 mm (Oval)	Porcine	Dried/Soft	0,4-0,6 mm	About 5 months
LS25FS	25x25 mm	Porcine	Dried/Soft	0,4-0,6 mm	About 5 months
LS24LS	20x40 mm	Porcine	Dried/Soft	1 mm	About 6 months
LS03SS	30x30 mm	Porcine	Dried/Soft	2-4 mm	About 8 months
LS15LS	35x15 mm	Porcine	Dried/Hard	0,6-0,8 mm	About 7 months
LS35LS	35x35 mm	Porcine	Dried/Hard	1 mm	About 6 months
LS10HE	35x35 mm (Curved)	Equine	Dried/Soft	1 mm	About 6 months
LS25FE	25x25 mm	Equine	Dried/Soft	0,4-0,6 mm	About 5 months

**Lamina®**



## Membranes

Codes	Size	Origin	Type	Thickness	Mean observed re-entry time
EM03XS	30x30 mm	Porcine	Dried	X-Fine	About 2 months
EM02HS	20x20 mm	Porcine	Dried	Standard	About 4/6 months
EM03HS	30x30 mm	Porcine	Dried	Standard	About 4/6 months
EV02LLE	20x20 mm	Equine	Dried	Fine	About 3/4 months
EV03LLE	30x30 mm	Equine	Dried	Fine	About 3/4 months
EV02HHE	20x20 mm	Equine	Dried	Standard	About 4/6 months
EV03HHE	30x30 mm	Equine	Dried	Standard	About 4/6 months
EV04LLE	40x40 mm	Equine	Dried	Fine	About 3/4 months
EV06LLE	80x60 mm	Equine	Dried	Fine	About 3/4 months

**Evolution**



Codes	Size	Origin	Type	Thickness	Mean observed re-entry time
ED02LS	20x20 mm	Porcine	Dried	0,4-0,6 mm	About 1 month
ED75SS	7x5 mm	Porcine	Dried	2-3 mm	About 5 months
ED15SS	15x5 mm	Porcine	Dried	2-3 mm	About 5 months
ED03SS	30x30 mm	Porcine	Dried	2-3 mm	About 5 months
ED05FS	50x50 mm	Porcine	Dried	0,8-1 mm	About 3 months
ED21FS	12x8 mm (Oval)	Porcine	Dried	0,8-1 mm	About 3 months
ED25FS	25x25 mm	Porcine	Dried	0,8-1 mm	About 3 months
ED05SS	50x50 mm	Porcine	Dried	2-3 mm	About 5 months

**Derma**



# OsteoBiol®

by Tecnoss

Made in Italy

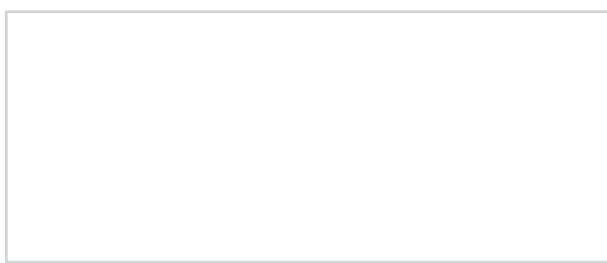


[o s t e o b i o l . c o m](http://osteobiol.com)

**Tecnoss® s.r.l.**  
info@tecnoss.com  
www.tecnoss.com

BIOMATERIALS ENGINEERING

DISTRIBUTED BY



**Tecnoss® Dental s.r.l.**  
info@tecnoss-dental.com  
www.osteobiol.com

INTERNATIONAL  
SALES & MARKETING